

# Portuguese Pancreatic Club Perspectives on Pancreatic Neuroendocrine Neoplasms: Diagnosis and Staging, Associated Genetic Syndromes and Particularities of Their Clinical Approach

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## Keywords

Pancreatic neuroendocrine neoplasm · Diagnosis · Staging · Multiple neuroendocrine neoplasia type 1 · Von Hippel-Lindau disease

## Abstract

Pancreatic neuroendocrine neoplasms (panNENs) have been historically regarded as rare, but their incidence has raised more than 6-fold over the last 3 decades, mostly owing to improvement in the detection of small asymptomatic tumours with imaging. Early detection and proper classification and staging are essential for the prognosis and management of panNENs. Histological evaluation is mandatory in all patients for the diagnosis of panNEN. Regarding localization and staging, multiphasic contrast-enhanced computer tomography is considered the imaging study of

choice. Nevertheless, several other diagnostic modalities might present complementary information that can help in diagnosis and staging optimization: magnetic resonance imaging, somatostatin receptor imaging using positron emission tomography in combination with computed tomography (PET/CT), PET/CT with fluorodeoxyglucose (<sup>18</sup>F-FDG), and endoscopic ultrasound. Approximately 10% of panNENs are due to an inherited syndrome, which includes multiple endocrine neoplasia type 1, von Hippel-Lindau disease, neurofibromatosis type 1 (NF-1), tuberous sclerosis complex, and Mahvash disease. In this review, the Portuguese Pancreatic Club summarizes the classification, diagnosis, and staging of panNENs, with a focus on imaging studies. It also summarizes the characteristics and particularities of panNENs associated with inherited syndromes.

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**Perspetivas do Clube Português do Pâncreas sobre Neoplasias Neuroendócrinas Pancreáticas: Diagnóstico e Estadiamento, Síndromes Genéticas Associadas e Particularidades da sua Abordagem Clínica**

**Palavras Chave**

Neoplasia neuroendócrina pancreática · Diagnóstico · Estadiamento · Neoplasia neuroendócrina múltipla tipo 1 · Doença de Von Hippel-Lindau

**Resumo**

As neoplasias neuroendócrinas pancreáticas (panNENs) são historicamente consideradas raras, embora a sua incidência tenha aumentado mais de 6 vezes nas últimas três décadas, principalmente devido à otimização do diagnóstico de tumores pequenos e assintomáticos em exames de imagem. A deteção precoce, a classificação e o estadiamento adequados são essenciais para o prognóstico e abordagem dos panNENs. A avaliação histológica é obrigatória em todos os doentes para o diagnóstico de panNENs. Para a localização e estadiamento, a TC multifásica com contraste é considerada o estudo de imagem de eleição. Contudo, várias outras modalidades diagnósticas podem apresentar informações complementares que podem auxiliar no diagnóstico e na otimização do estadiamento: ressonância magnética, PET/CT dos receptores da somatostatina, PET/CT [<sup>18</sup>F]FDG e ecoendoscopia. Aproximadamente 10% dos panNENs estão relacionados com síndromes hereditários, que incluem neoplasia endócrina múltipla tipo 1 (MEN1), doença de von Hippel-Lindau (VHL), neurofibromatose tipo 1 (NF1), complexo de esclerose tuberosa (TSC) e doença de Mahvash. Neste artigo, o Clube Português de Pâncreas aborda a classificação, diagnóstico e estadiamento de panNENs, com foco nos estudos de imagem, bem como resume as características e particularidades dos panNENs associados aos síndromes hereditários.

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**Introduction**

Neuroendocrine neoplasms (NENs) can arise from different organs, even though the lungs, gastrointestinal (GI) tract, and pancreas are the most common sites. GI

and pancreatic NENs (panNENs) are histologically classified under the same category; however, panNENs have particular clinical features from GI NENs and, thus, should be considered separately. PanNENs have been historically regarded as rare, but the reported prevalence in autopsy series (0.8–10%) is much higher than in population-based studies and their incidence has raised more than 6-fold over the last 3 decades [1]. This growth occurred across all disease stages and tumour grades, but it was particularly pronounced for localized low-grade tumours due to the widespread use of advanced imaging tests [1].

PanNENs are divided into functional and non-functional tumours. About 70–90% of panNENs are classified as non-functional [2]. Functional tumours secrete particular hormones or peptides, such as insulin, gastrin, vasoactive intestinal peptide, glucagon, and somatostatin.

Inherited syndromes are important to recognize in the setting of panNENs, as they have significant implications for patient's medical management, representing an opportunity for early detection of subsequent manifestations. Although most panNENs are sporadic, approximately 10% are due to an inherited syndrome, most commonly multiple endocrine neoplasia type 1 (MEN1) and, less commonly, von Hippel-Lindau disease (VHL), neurofibromatosis type 1 (NF-1), or tuberous sclerosis complex (TSC) [3]. Mahvash disease, a recently described inherited disease, has also been associated with panNENs.

In this review, the Portuguese Pancreatic Club summarizes the classification, diagnosis, and staging of panNENs, with a focus on imaging studies. It also summarizes the characteristics and particularities of the diagnosis and treatment of panNENs associated with inherited syndromes.

**PanNEN Diagnosis**

In accordance with World Health Organization (WHO) criteria, panNEN definitive diagnosis requires histological confirmation [4]. For appropriate pathological diagnosis, morphology, grading and immunohistochemical staining for chromogranin A (CgA) and synaptophysin should be reported. The neuroendocrine phenotype is confirmed by the immunohistochemical detection of one of the two neuroendocrine markers commonly used: synaptophysin and CgA. Other neuroendocrine markers, such as neuron-specific enolase and CD56, can be positive in panNEN, but lack specificity [5].

According to WHO 2019 classification, NENs are classified according to morphology and proliferation (Ki67 index and mitotic count) into well-differentiated

**Table 1.** WHO 2019 classification for gastroenteropancreatic NENs [4]

Morphology	Grade	Mitotic count, 2 mm <sup>2</sup>	Ki67 index, %
Well-differentiated NETs	G1	<2	<3
Well-differentiated NETs	G2	2–20	3–20
Well-differentiated NETs	G3	>20	>20
Poorly differentiated NECs	G3	>20	>20
Small-cell			
Large-cell			

NEC, neuroendocrine carcinoma; NEN, neuroendocrine neoplasm; NET, neuroendocrine tumour; WHO, World Health Organization.

NENs (G1 to G3) and poorly differentiated neuroendocrine carcinomas (NEC) (always G3) (Table 1) [4]. If Ki67 index and mitotic count are discordant, the higher grade is attributed [6]. Well-differentiated NENs and NEC are biologically and genetically two different diseases. Furthermore, clear prognostic differences can be seen between these two conditions, even in grade 3 NEN. Therefore, the most recent WHO classification split the heterogeneous G3 NEN into well-differentiated NEN G3 and poorly differentiated NEC G3 [4, 6].

Clinical suspicion of functional panNENs can be confirmed by the measurement of specific hormones secreted by functional tumours (i.e., insulin, proinsulin, glucagon, gastrin, vasoactive intestinal polypeptide) and its correlation with hormonal symptoms [5]. Hormone levels also correspond to changes in tumour burden and can therefore serve as specific tumour markers during follow-up. If insulinoma is suspected, serum level of insulin and C-peptide along with glucose during prolonged fasting (72 h) is useful for the diagnosis [7]. Patients with insulinoma present abnormally elevated levels of insulin and C-peptide during hypoglycemia. Proinsulin levels are also elevated in insulinoma. When gastrinoma is suspected, fasting serum gastrin levels should be evaluated. A fasting serum gastrin level that is 10 times greater than the upper limit of the normal range along with a gastric pH <2 is diagnostic of gastrinoma [8]. Demonstration of gastric acid hypersecretion by means of gastric fluid pH evaluation in particular can be obtained through nasogastric tube or upper endoscopy using electrode, filter paper, or biochemical evaluation of aspirate [9]. However, given the advent of highly sensitive imaging-techniques and problems with the classical diagnosis of gastrinomas (intermediate levels of fasting serum gastrin and widespread use of proton pump inhibitors) most expert centres have adopted an alternative diagnostic work-up that include upper endoscopy, endoscopic ultrasonography, magnetic resonance imaging (MRI), and

somatostatin receptor (STTR) imaging [9]. Serologic CgA has a limited specificity for the diagnosis of panNEN; however, it might be useful in follow-up [10]. Immunohistochemistry staining for peptide hormones such as gastrin, insulin and glucagon can also help confirm the source of a clinical symptomatology, but there is no complete agreement between immunohistochemistry and symptomatology [11]. Approximately 10% of non-functional panNENs are multiple (multifocal) [10].

### PanNEN Staging

Disease stage and tumour grade are the two major independent prognostic parameters and should always be assessed to select the best therapeutic strategy. Tumour size >2 cm and Ki67 index >3% are predictors of metastatic disease, which is associated to decreased survival. Regarding panNENs staging, the tumour, node, and metastasis staging system proposed by the European Neuroendocrine Tumour Society (ENETS) and adopted in the eighth edition of the American Joint Committee on Cancer (AJCC) is recommended (Table 2) [12]. For NECs, however, the staging system of adenocarcinomas should be applied [12]. Furthermore, the primary tumour site has also an impact on the prognosis in advanced disease and should be detected in the presence of metastatic NEN. In fact, data show that patients with metastatic panNENs have a less favourable prognosis than patients with metastatic small intestinal NENs [13].

Computed tomography (CT) is the most widely used method for NEN imaging because of its availability and high diagnostic yield [14]. The ENETS, North American Neuroendocrine Tumour Society (NANETS), and European Society for Medical Oncology (ESMO) consensus guidelines recommend pancreatic protocol CT (three-phase CT) as the best imaging modality for staging, primarily due to the characterization of vascular

**Table 2.** Neuroendocrine tumours of the pancreas, TNM staging AJCC UICC 8th edition [11]

Primary tumour (T)			
T category	T criteria		
Tx	Tumour cannot be assessed		
T1	Tumour limited to the pancreas, <2 cm		
T2	Tumour limited to the pancreas, 2–4 cm		
T3	Tumour limited to the pancreas, >4 cm; or tumour invading the duodenum or common bile duct		
T4	Tumour invading adjacent organs (stomach, spleen, colon, adrenal gland) or the wall of large vessels (celiac axis or the superior mesenteric artery)		
Regional lymph nodes (N)			
N category	N criteria		
NX	Regional lymph nodes cannot be assessed		
N0	No regional lymph node involvement		
N1	Regional lymph node involvement		
Distant metastasis (M)			
M category	M criteria		
M0	No distant metastasis		
M1	Distant metastases		
M1a	Metastasis confined to liver		
M1b	Metastases in at least one extrahepatic site (e.g., lung, ovary, nonregional lymph node, peritoneum, bone)		
M1c	Both hepatic and extrahepatic metastases		
Prognostic stage groups			
when T is . . .	and N is . . .	and M is . . .	then the stage group is . . .
T1	N0	M0	I
T2	N0	M0	II
T3	N0	M0	II
T4	N0	M0	III
Any T	N1	M0	III
Any T	Any N	M1	IV

TNM, tumour node metastasis; AJCC, American Joint Committee on Cancer; UICC, Union for International Cancer Control.

involvement of the primary tumour. The sensitivity of CT to detect panNENs is 61–93% and the specificity is 71–100% [14–16]. The detection rate for liver metastases (LMs) is 79% [17, 18], and for extra-abdominal soft tissue metastases is 70% [19]. However, despite the good diagnostic performance of CT, this radiological method shows lower than desired sensitivity for the detection of small metastatic lymph nodes (<1 cm), bone metastases, and small peritoneal metastases [20].

MRI is a relevant complementary imaging test in the staging of patients with panNENs. In fact, abdominopelvic MRI protocol is advantageous for examination

of the liver and the pancreas, when compared with CT, and is also recommended in the initial staging, as it outperforms CT for imaging small LMs [11]. Currently, diffusion-weighted imaging with MRI as well as magnetic resonance cholangiopancreatography is routinely applied, facilitating lesion detection. The MRI sensitivity to detect panNENs is described as 54–100%. The sensitivity of MRI for detection of LMs is 82–98% [21]. MRI is also superior to CT for imaging of the bones and the brain. However, MRI may miss small lung metastases, and CT is preferred for imaging of the lungs [14]. Despite this, MRI protocols, similar to CT

protocols, are usually restricted to images from the 11th vertebra body through the iliac crest in the staging of panNENs.

Endoscopic ultrasound (EUS) is the current optimal imaging method to diagnose small panNENs, being able to detect lesions as small as 2–3 mm in diameter. EUS is reported as having 82–93% sensitivity and 86–95% specificity for the detection of panNENs [22]. When compared with CT, EUS appears to have a better diagnostic performance, being able to detect lesions not apparent in other diagnostic modalities [23]. In the ENETS consensus guidelines, EUS is the imaging study of choice when other non-invasive imaging studies are negative, allowing screening the entire pancreas and a detailed evaluation of the tumour [5]. EUS has been shown to be superior to CT in the detection and localization of panNENs in patients with MEN1 syndrome, where the tumours are usually small and multifocal [24]. Furthermore, contrast-enhanced EUS (CE-EUS) is helpful in characterizing small panNENs, which are incidentally found on other imaging modalities [25]. Over 90% of panNENs showed hypoechogenicity in B-mode and hyperechogenicity after the injection of contrast agent in contrast-enhanced EUS and up to 75% of hypervascular lesions on CE-EUS were NENs in a recent study [26]. Another benefit of EUS is that it allows for tissue acquisition, using fine-needle aspiration for cytology or, fine-needle biopsy (FNB) with a cutting needle for histopathological diagnosis. Several studies have documented better diagnostic performance with end-cutting FNB needles, particularly for Ki67 index determination (tumour grading), which may be underestimated in fine-needle aspiration samples [27, 28]. Preoperative knowledge of tumour grading is relevant for treatment decision, particularly in small (<2 cm) panNENs, for which tumour grading should be considered in the choice between surgery and surveillance [10, 26].

When diagnosing most NENs, STTRs imaging with positron emission tomography in combination with CT (PET/CT) using radiolabelled somatostatin analogues (e.g., [<sup>68</sup>Ga]DOTATOC, [<sup>68</sup>Ga]DOTANOC, [<sup>68</sup>Ga]DOTATATE) is highly sensitive and should be included in the tumour staging process [14]. It offers a high detection rate of lymph node, bone, and peritoneal lesions, as well as a high detection rate of primary lesions in patients with unknown primary tumours [11]. In the past, when PET/CT was not accessible, STTR scintigraphy (OctreoScan™) was used, with significantly less sensitivity [11]. The sensitivity to detect panNEN by STTR-PET/CT is 70.5–87.0%, and the specificity is 75–100% [29]. For the detection of bone metastases, STTR-PET/CT shows a sensitivity of 97–100% and a specificity of 92–100% [30]. Notwithstanding, the potential for false-positive uptakes must be considered,

particularly within the uncinate process and the pancreatic head [28]. STTR-PET/CT should also not be used in the differential diagnosis between panNENs and other hypervascular nodules, such as ectopic spleen – for characterizing pancreatic nodules detected on MRI or CT, EUS with biopsy is superior [10].

[<sup>18</sup>F]FDG-PET/CT is an additional diagnostic tool for patients with panNENs, particularly those equal to or higher than G2, characterized by higher glucose metabolism and lower STTR expression than low-grade NENs [31]. Furthermore, it provides prognostic value since FDG-positive NEN lesions associate with a worse prognosis [31, 32]. Combined STTR and FDG-PET/CT imaging (dual tracer PET/CT) have shown complementary lesion detection [31]. However, the benefits of this combination are not validated and should only be adopted on an individual basis, balancing the potential advantages with the increasing costs [11]. Regarding diagnosis and staging of NECs, FDG-PET/CT is of central importance, since STTR-PET/CT has low sensitivity.

Other diagnostic techniques such as contrast-enhanced ultrasound (US) and intraoperative US might be useful in the localization and staging of panNENs [11]. In fact, intraoperative US has an excellent diagnostic performance for the detection of lesions located in the pancreas and liver and is mandatory before pancreatic resection in MEN1 syndrome patients [11].

In summary, disease staging is a major independent prognostic parameter and should always be assessed. CT or MRI protocols with abdominopelvic imaging should be performed with additional anatomical segments evaluation in case of findings suggestive of metastatic disease. Based on current evidence, MRI should be preferred for the detection of the liver, pancreas, brain, and bone lesions, while CT is preferred for imaging of the lungs. Whole-body STTR-PET/CT imaging is complementary to CT or MRI and should also be part of the tumour staging. A summary of the imaging staging recommendations from different international societies can be found in Table 3.

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### Genetic Syndromes Associated with panNENs and Particularities of Their Clinical Approach

Five different inherited syndromes are associated with the development of panNENs: MEN1, VHL, NF-1, also known as Von Recklinghausen disease, TSC (Table 4), and the more recently described Mahvash disease. Mahvash disease (the rarest among the five known hereditary panNEN syndromes) is the only recessively inherited PNET syndrome (associated with a mutation in



**Table 3.** Imaging staging recommendations from international societies

ESMO	ENETS	NANETS	JNETS
<p>CT/MRI</p> <ul style="list-style-type: none"> <li>• MRI is preferred to CT for the detection of the liver, pancreas, brain, and bone lesions</li> <li>• CT is preferred for imaging of the lungs</li> </ul>	<ul style="list-style-type: none"> <li>• CT/MRI (including MRCP) is recommended</li> <li>• The decision between CT or MRI may depend on the expertise of the institution</li> </ul>	<ul style="list-style-type: none"> <li>• CT/MRI are excellent tools for evaluating primary tumours and nodal metastases</li> <li>• MRI is better than CT for imaging hepatic metastases</li> </ul>	<ul style="list-style-type: none"> <li>• CT/MRI is recommended</li> </ul>
<p>PET/CT</p> <ul style="list-style-type: none"> <li>• 68Ga/64Cu PET/CT imaging should be part of the tumour staging</li> <li>• FDG-PET/CT is optional in NENs</li> </ul>	<ul style="list-style-type: none"> <li>• 68 Ga PET/CT using is recommended for extrahepatic disease manifestation</li> </ul>	<ul style="list-style-type: none"> <li>• 68Ga/68Ga PET/CT is recommended for identifying primary tumours and the extent of metastatic disease</li> </ul>	<ul style="list-style-type: none"> <li>• 68Ga/68Ga PET/CT is recommended as it offers high specificity</li> </ul>
<p>EUS</p> <ul style="list-style-type: none"> <li>• Not referred</li> </ul>	<ul style="list-style-type: none"> <li>• Small NF pancreatic NETs may be better assessed using EUS</li> <li>• EUS-FNB has good results in confirming a diagnosis</li> </ul>	<ul style="list-style-type: none"> <li>• EUS-FNB should be performed when histologic diagnosis of a panNEN would be helpful or when there is a question about tumour grade</li> </ul>	<ul style="list-style-type: none"> <li>• EUS-FNB is recommended when performing histology is recommended</li> </ul>

ESMO, European Society for Medical Oncology; ENETS, European Neuroendocrine Tumour Society; NANETS, North American Neuroendocrine Tumour Society; JNETS, Japan Neuroendocrine Tumour Society; CT, computed tomography; MRI, magnetic resonance imaging; PET/CT, positron emission tomography in combination with CT; EUS, endoscopic ultrasound; FDG, fluorodeoxyglucose; MRCP, magnetic resonance cholangiopancreatography; FNB, fine-needle biopsy; panNEN, pancreatic neuroendocrine neoplasia.

**Table 4.** Characterization of geNENic syndromes associated with panNEN

Syndrome	Prevalence (per/100,000 population)	Frequency of panNENs	Type of panNEN
MEN1	1–10	20–80%	NF-panNENs Gastrinoma insulinoma Glucagonoma vipoma GRFoma Somatostatinoma
VHL	2–3	10–17%	NF-panNENs
NF-1	20–25,000	0–10%	Duodenal somatostatinoma
TSC	10	Uncommon	Gastrinoma, insulinoma NF-panNENs
MD	11 cases described	100%	NF-panNENs

MEN1, multiple endocrine neoplasia type 1; VHL, Von Hippel-Lindau Disease; NF-1, neurofibromatosis 1; TSC, tuberous sclerosis complex; MD, Mahvash disease; panNEN, pancreatic neuroendocrine neoplasia; NF-panNEN, non-functional panNEN; GRF, growth hormone-releasing factor.

the glucagon receptor gene), has a penetrance of 100% and appears to be exclusively associated with the development of non-functional panNENs. So far, only 11 cases have been describe in the literature, however, the

clinical impact of Mahvash disease on panNENs is likely higher than that of NF-1 and TSC, which have very low panNEN penetrance [33]. The first four hereditary panNEN syndromes will be further discussed below.

### Multiple Endocrine Neoplasia Type 1

MEN1 is a rare, autosomal-dominant, syndrome secondary to mutations in the *MEN1* gene on chromosome 11q13, which encodes the protein menin. Menin has an important role in regulating cell growth, cell cycle progression, and various other cellular processes [34]. Classically, MEN1 is characterized by the development of tumours/hyperplasia in multiple endocrine tissues (parathyroid, pancreas, and pituitary); however, other tumours can also be associated with MEN1, including the adrenal, skin, thyroid, CNS, smooth muscles as well as carcinoid (lung and thymus) tumours and gastric NENs [34–38].

As much as 20–80% of all patients with MEN1 will develop clinically relevant panNENs (Table 4) [34]. MEN1 occurs in 20–30% of all patients with gastrinomas and Zollinger-Ellison syndrome (ZES), 5% of patients with insulinomas, and <3% with non-functional panNENs [34]. In other types of functional panNENs, known as rare functioning tumours, MEN1 is the most frequent familial condition associated, with glucagonomas occurring in 3% of MEN1 patients, VIPomas in 3%, and GRHomas and somatostatinomas in <1% [39].

Non-functional panNENs are among the most common tumours of the pancreaticoduodenal region in patients with MEN1, with a penetrance as high as 35% at 50 years of age. In most patients, non-functional panNENs are  $\leq 2$  cm and therefore the risk of metastasis and death is very low [34]. However, average life expectancy for patients with non-functional panNENs is similar to that for patients with gastrinomas and shorter than that for patients without panNENs.

The best way to diagnose and stage non-functional panNENs in patients with MEN1 is unclear. Assays for tumour markers like CgA have low value [40]. Similar to panNENs not associated with MEN1, imaging modalities appear to be ideal for the diagnostic work-up of these patients as well as for the screening of panNENs in patients with MEN1. Current guidelines recommend annual imaging with CT, MRI, or EUS for the screening of panNENs in patients with MEN1 starting at the age of 18 years old [41]. However, given the low growth rate of these lesions, 2–3 years intervals may be considered in patients with previously negative surveillance. Recent data suggest that EUS outperforms CT scanning in this setting, and a combination of MRI plus EUS has been recommended [42]. Furthermore, STTR-PET/CT scanning has been reported to have high sensitivity for panNEN in MEN1, often leading to a change in management [43]. However, high radiation dose has been reported during surveillance program in patients with MEN1 [44]. As such, multi-

modality imaging strategies designed to minimize radiation exposure should be considered.

The management of non-functional panNENs in MEN1 is controversial [45]. Many experienced centres have been using a tumour size threshold of 2 cm in the decision to surgically resect non-functional panNEN [46], with subsequent yearly surveillance [47]. Generally speaking, tumours under 1 cm have a low risk for substantial growth and metastasis and avoiding surgery with continued surveillance is reasonable. Available data for tumours between 1 and 2 cm are less clear. Triponez et al. demonstrated that the risk of death was low for patients with panNEN <2 cm and proposed a conservative attitude for these patients in the absence of aggressive features such as rapid progression on imaging studies [48].

In patients with MEN1, panNENs are responsible for 30% of the ZES, with duodenal NENs being responsible for the remainder. Most gastrinomas are well differentiated, with a low proliferative activity (Ki67 index), usually close to 2%. Immunohistochemically, almost all gastrinomas stain for gastrin [49]. Patients with ZES and MEN1 present at an earlier age (mean 32–35 years) than patients with sporadic disease [34]. Of all MEN1/ZES patients, 25% lack a family history of MEN1, supporting the need to suspect of MEN1 in all patients with ZES [34]. In up to 45% of MEN1 patients, the symptoms of ZES precede those of hyperparathyroidism and can be the initial symptoms [50]. However, almost all MEN1 patients have hyperparathyroidism at the time of ZES diagnosis, although in many patients it can be asymptomatic [34, 50]. Regarding insulinomas, approximately 5% are associated with MEN1 syndrome. Comparative studies between patients with insulinomas, with or without MEN1 are lacking. Finally, the association between MEN1 and rare functioning tumours is less clear.

Because of the frequent association between MEN1 and ZES, and in similarity to patients with a clinical diagnose of MEN1 (Table 5), all patients with ZES should have biochemical studies for MEN1. Similarly, all patients with insulinoma or rare functioning tumours with suspicion of MEN1 (Table 5) should have the same evaluation. Serum parathormone levels, ionized calcium levels and prolactin levels should be performed at initial evaluation and during yearly follow-up in patients with ZES [34, 50]. Furthermore, all patients suspected of MEN1 (Table 5) need to be assessed for the other tumours, which are generally non-functional [49]. Specific parathyroid studies are required if hyperparathyroidism is found (US, CT/MRI, 99m Tc-sestamibi scan) [51]. All patients require MRI of the *sella turcica* region and, after 20 years of age, require CT of the chest/abdomen [52]. If MEN1/ZES is

**Table 5.** MEN1 genetic testing criteria

Individuals meeting any one of the following should be referred for genetic counselling and gene testing

- Patient meeting MEN1 clinical diagnostic criteria:
  - 1) Two or more classic MEN1-associated tumours (parathyroid adenoma, pituitary adenomas, or GEP-NEN)
  - 2) Single MEN1-associated tumour and a first-degree relative with MEN1
    - First-degree relative of a patient with MEN1 syndrome
    - Parathyroid adenoma diagnosed before age 30
    - Multiple parathyroid adenomas
    - Gastrinoma or multiple panNEN at any age
    - Single panNEN diagnosed before age 20
    - Female with thymic NEN
    - One classic MEN1-associated tumour and one nonclassic feature (carcinoid tumour, dermatologic features, or adrenal tumour)

MEN1, multiple endocrine neoplasm type 1; NEN, neuroendocrine neoplasia.

present, UGI endoscopy for gastric NEN is recommended [53]. Routine SRS is not recommended if other imaging studies for NEN are negative. EUS is more sensitive than cross sectional imaging studies (CT, MRI, US) for the detection and characterization of small non-functional panNENs. However, since routine surgical resection of small panNENs (<2 cm) is not recommended and the EUS criteria on when to operate these patients are not established the added benefit of EUS is controversial [54].

Regarding genetic testing, it should be performed in patients meeting criteria for MEN1 ( $\geq 2$  tumours associated with MEN1), patients with gastrinomas or patients meeting further criteria described in Table 5. If genetic testing is considered, genetic counselling should be offered, prior to testing [51, 55].

In MEN1/ZES patients, surgery without a Whipple resection is associated with >90% of recurrence [34, 49, 56]. Therefore, routine surgical exploration is controversial in patients with MEN1/ZES. Indeed, these patients usually have multiple gastrinomas, frequently with lymph node metastases, with concomitant panNENs (non-functional primarily), are rarely cured and have an excellent life expectancy if only small tumours (<2 cm) are present [34, 49, 57]. However, surgery is the only treatment approach with curative intent [56]. As such, it has been generally recommended that surgery should be performed in patients with MEN1 and panNENs >2 cm [48]. In patients with MEN1 and insulinoma, in which multiple tumours are frequently present, the aim of surgery is to control inappropriate insulin secretion by excising all insulinomas. As such, preoperative

localization of which pancreatic tumours are the insulinomas is mandatory because these patients frequently have other panNENs (usually non-functional) [58].

The prognostic significance of MEN1 in patients with panNENs is not entirely clear. Some studies in patients with gastrinomas suggest these patients have a better prognosis, even though the gastrinomas are almost always multiple [59]. With the ability to treat both the ZES and the hyperparathyroidism, recent studies show that in patients with MEN1, the natural history of the panNEN is increasingly becoming a determinant of survival [34]. Finally, patients with MEN1 frequently have multiple insulinomas, however, these are usually cured surgically [60]. There are no comparative studies on survival in MEN1 patients with insulinomas compared to sporadic cases.

#### *Von Hippel-Lindau Disease*

VHL is a rare autosomal-dominant disease caused by mutations in the *VHL* gene, on chromosome 3p25, that encodes the peptide pVHL, important in the regulation of angiogenic growth and the activity of several mitotic factors (VEGF, PDGF, TGF $\alpha$ , erythropoietin) [34]. VHL is characterized by hemangioblastomas of the retina and cranio-spinal region, endolymphatic sac tumours, renal cell carcinomas or cysts, pheochromocytomas, and epididymal cystadenomas (Table 6). Furthermore, pancreatic tumours or cysts can be present in 35–77% of patients [46, 59]. Specifically, panNENs develop in 10–17% of patients with VHL, and in almost all cases they are non-functional panNENs, and usually are asymptomatic (Table 4) [61]. In contrast to MEN1 patients, most VHL patients have a single panNEN, although patients might also present with multifocal lesions [34]. The majority of panNENs in VHL are well differentiated (grade 1 or 2), small (<2 cm) and present a slow growth when compared with sporadic tumours. As a result of its clinical indolence, many of these lesions are diagnosed incidentally during routine VHL surveillance for renal lesions [62]. However, in 8–50% of VHL patients, panNENs are metastatic and LMs occur in 9–37% [63]. In patients with VHL, panNENs are more likely to metastasize when present with size >3 cm in diameter, rapid tumour doubling time (<500 days), and VHL missense and/or exon 3 pathogenic variants [64]. Based on these 3 risk factors, risk stratification of panNENs in the context of VHL has been suggested for management optimization [64]. As such, in the presence of a panNEN, if clinical features suggest VHL (Table 6), appropriate gene testing should be considered after genetic counseling [34], and molecular imaging, typically with STTR-PET/CT should be offered [65].



**Table 6.** Von Hippel-Lindau disease (VHL) diagnostic criteria

Simplex case (no family history), individual with $\geq 2$ of the following
<ul style="list-style-type: none"><li>• Two or more hemangioblastomas of the retina, spine, or brain or a single hemangioblastoma in association with a visceral manifestation (e.g., multiple kidney or pancreatic cysts)</li><li>• Renal cell carcinoma</li><li>• Paraganglioma or pheochromocytoma</li><li>• Less commonly, endolymphatic sac tumours, papillary cystadenomas of the epididymis or broad ligament, or neuroendocrine tumours of the pancreas</li></ul>
Familial case (known relative with VHL), individual with $\geq 1$ of the following
<ul style="list-style-type: none"><li>• Retinal angioma</li><li>• Spinal or cerebellar hemangioblastoma</li><li>• Paraganglioma or pheochromocytoma</li><li>• Renal cell carcinoma</li><li>• Multiple renal and pancreatic cysts</li></ul>

Management of panNENs is primarily surgical, although the criteria for surgical resection differ from those of patients with sporadic panNENs. Surgical resection should be reserved for patients with potentially resectable lesions greater than 3 cm in diameter in the body or tail of the pancreas, or greater than 2 cm in diameter in the head of the pancreas [65]. The hypoxia-inducible factor-2 $\alpha$  (HIF-2 $\alpha$ ) inhibitor Belzutifan, rather than other systemic therapies, is considered the best approach if surgery is not feasible, or the tumour is considered unresectable [65]. Nonoperative approaches (e.g., surveillance, Belzutifan) are appropriate for small primary lesions ( $\leq 3$  cm), incorporating other clinical factors such as type and location of the VHL pathogenic variant, and rate of tumour growth [63]. Regarding prognosis, VHL-associated panNENs tend to be associated with a more indolent course and, similarly, long-term outcomes of resected VHL-associated panNENs appear to be generally better than those of sporadic [66].

#### *Neurofibromatosis Type 1*

NF-1 is an autosomal-dominant syndrome with a population frequency of 1 in 3,000 births, with half of cases due to a de novo mutation [67]. NF-1 is due to mutations in the *NF-1* gene on chromosome 17q11.2, which encodes the protein neurofibromin, which affects cell growth, through Ras protein activation and mammalian target of rapamycin (mTOR) cascade regulation [46, 62]. This condition is characterized by the

development of several tumours, including neurofibroma, pheochromocytoma, and GI stromal tumour (Table 1) [46, 62]. CNS abnormalities are frequent with learning disorders (30–60%), attention deficit hyperactivity disorder, and epilepsy [46, 62]. Furthermore, panNENs occur only in a minority of NF-1 patients (10%) and are almost exclusively duodenal somatostatinomas (Table 4) [46, 63, 64]. However, NF-1 patients have been reported with NF-panNENs, ZES and insulinomas [34]. Duodenal somatostatinomas characteristically occur in the peri-ampullary region, have a mean size of 2.8 cm (range 1–5), comprise 23% of all ampullary NENs in several series, metastasize in 30% of cases, and are rarely associated with the clinical somatostatinoma syndrome (1–2%) [46]. To this date, there are no data to allow specific management recommendations of panNENs in the context of NF-1 and patients with NF-1 with panNENs are usually treated as sporadic panNENs. However, it is important to remember that in patients with panNENs, if clinical features suggest NF-1, appropriate gene testing should be considered after genetic counseling [34].

#### *Tuberous Sclerosis Complex*

TSC is an autosomal-dominant disease caused by a mutation in one of two genes: the *TSC1* gene (which encodes hamartin) or the *TSC2* gene (which encodes tuberin). Both hamartin and tuberin are codependent and play a role in mTOR cascade regulation, protein translation, protein synthesis, and cell proliferation [34]. This condition is characterized by the development of hamartomas in multiple organs, neurological features (autism, mental retardation, epilepsy), and dermatological features (hypomelanotic macules, shagreen patches, ungual fibromas, facial angiofibromas) [68]. A small percentage of TSC patients (1%) have been reported to have panNENs, including gastrinomas, insulinomas, and non-functional panNENs, some of which are metastatic (Table 4) [69]. As such, in patients with panNENs, if clinical features suggest TSC, appropriate gene testing should be considered after genetic counselling [34]. However, to this date, specific management recommendations of panNENs in the context of tuberous sclerosis are lacking and patients are usually treated as sporadic ones.

#### **Conclusion**

Among pancreatic neoplasms, panNENs are rare tumours. However, their incidence is increasing with the advancement of imaging technology and increased

opportunities for checking pancreatic diseases. Early detection and proper staging are paramount for prognostication and for treatment decision. Histological evaluation is mandatory in all patients for the diagnosis of panNENs and plays a central role in grading these tumours, essential for proper prognostic information.

Regarding localization and staging, multiphasic contrast-enhanced CT is considered the imaging study of choice. MRI has been suggested to be more sensitive in detecting small tumours and LM than other modalities. STTR-PET/CT has been approved for the diagnosis and staging of panNENs and has improved sensitivity when compared with the more traditional STTR scintigraphy. EUS can detect tumours as small as 2–3 mm. Its sensitivity is equal or superior to multi-detector CT or MRI for their detection. Other benefits of EUS include the detection of lymph node involvement and vascular invasion and the possibility of tissue acquisition through FNB, which is crucial for tumour grading. EUS-FNB is the best modality for the differential diagnosis between panNENs and other hypervascular nodules detected on CT or MRI.

In the presence of inherited syndromes, panNENs may present with additional challenges. A thoughtful approach to the diagnosis and management is required, as these syndromes often involve multi-organ disease with a lifelong risk for tumour development. Additionally, the natural history of tumours in the setting of a hereditary condition may be different than it would be expected in a

sporadic form of the disease. The unique aspects to management, challenges in hereditary disease recognition and accurate diagnosis, and rarity of these syndromes should be kept present during the evaluation of patients with panNENs.

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### Conflict of Interest Statement

The authors have no conflicts of interest to declare.

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### Author Contributions

Joel Ferreira-Silva, Sara Meireles, Massimo Falconi, and Eduardo Rodrigues-Pinto were responsible for literature review and manuscript writing. Results were discussed in two meetings of Clube Português do Pâncreas and approved by all members. All authors critically reviewed and approved the final manuscript.

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### Reference

- 1 Lawrence B, Gustafsson BI, Chan A, Svejda B, Kidd M, Modlin IM. The epidemiology of gastroenteropancreatic neuroendocrine tumors. *Endocrinol Metab Clin North Am*. 2011;40(1):1–18, vii.
- 2 McKenna LR, Edil BH. Update on pancreatic neuroendocrine tumors. *Gland Surg*. 2014; 3(4):258–75.
- 3 Geurts JL. Inherited syndromes involving pancreatic neuroendocrine tumors. *J Gastrointest Oncol*. 2020;11(3):559–66.
- 4 Nagtegaal ID, Odze RD, Klimstra D, Paradis V, Rugge M, Schirmacher P, et al. The 2019 WHO classification of tumours of the digestive system. *Histopathology*. 2020;76(2):182–8.
- 5 Perren A, Couvelard A, Scoazec JY, Costa F, Borbath I, Delle Fave G, et al. ENETS consensus guidelines for the standards of care in neuroendocrine tumors: pathology: diagnosis and prognostic stratification. *Neuroendocrinology*. 2017;105(3):196–200.
- 6 Klöppel G, Klimstra DS, Hruban RH, Adsay V, Capella C, Couvelard A, et al. Pancreatic neuroendocrine tumors: update on the new World Health Organization classification. *AJSP: Rev Rep*. 2017;22(5): 233–9.
- 7 Vezzosi D, Bennet A, Fauvel J, Caron P. Insulin, C-peptide and proinsulin for the biochemical diagnosis of hypoglycaemia related to endogenous hyperinsulinism. *Eur J Endocrinol*. 2007;157(1):75–83.
- 8 Metz DC, Jensen RT. Gastrointestinal neuroendocrine tumors: pancreatic endocrine tumors. *Gastroenterology*. 2008;135(5): 1469–92.
- 9 Sorbye H, Grande E, Pavel M, Tesselaar M, Fazio N, Reed NS, et al. European Neuroendocrine Tumor Society (ENETS) 2023 guidance paper for digestive neuroendocrine carcinoma. *J Neuroendocrinol*. 2023;35(3):e13249.
- 10 Nehar D, Lombard-Bohas C, Olivieri S, Claustrat B, Chayvialle JA, Penes MC, et al. Interest of Chromogranin A for diagnosis and follow-up of endocrine tumours. *Clin Endocrinol*. 2004;60(5):644–52.
- 11 Pavel M, Öberg K, Falconi M, Krenning EP, Sundin A, Perren A, et al. Gastroenteropancreatic neuroendocrine neoplasms: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2020;31(7):844–60.
- 12 Brierley JD, Gospodarowicz MK, Wittekind C. TNM classification of malignant tumours. John Wiley & Sons; 2017.
- 13 Dasari A, Shen C, Halperin D, Zhao B, Zhou S, Xu Y, et al. Trends in the incidence, prevalence, and survival outcomes in patients with neuroendocrine tumors in the United States. *JAMA Oncol*. 2017;3(10):1335–42.
- 14 Sundin A, Arnold R, Baudin E, Cwikla JB, Eriksson B, Fanti S, et al. ENETS consensus guidelines for the standards of care in neuroendocrine tumors: radiological, nuclear medicine & hybrid imaging. *Neuroendocrinology*. 2017;105(3):212–44.
- 15 Gabriel M, Decristoforo C, Kendler D, Dobrozemsky G, Heute D, Uprimny C, et al. 68Ga-DOTA-Tyr3-octreotide PET in neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and CT. *J Nucl Med*. 2007;48(4):508–18.

- 16 Procacci C, Carbogogni G, Accordini S, Biasiutti C, Bicego E, Romano L, et al. Non-functioning endocrine tumors of the pancreas: possibilities of spiral CT characterization. *Eur Radiol*. 2001;11(7):1175–83.
- 17 Fidler JL, Fletcher JG, Reading CC, Andrews JC, Thompson GB, Grant CS, et al. Preoperative detection of pancreatic insulinomas on multiphasic helical CT. *AJR Am J Roentgenol*. 2003;181(3):775–80.
- 18 Gouya H, Vignaux O, Augui J, Dousset B, Palazzo L, Louvel A, et al. CT, endoscopic sonography, and a combined protocol for preoperative evaluation of pancreatic insulinomas. *AJR Am J Roentgenol*. 2003;181(4):987–92.
- 19 Kim JH, Eun HW, Kim YJ, Lee JM, Han JK, Choi BI. Pancreatic Neuroendocrine Tumour (PNET): staging accuracy of MDCT and its diagnostic performance for the differentiation of PNET with uncommon CT findings from pancreatic adenocarcinoma. *Eur Radiol*. 2016;26(5):1338–47.
- 20 Norlén O, Montan H, Hellman P, Stålborg P, Sundin A. Preoperative (68)Ga-DOTA-Somatostatin analog-PET/CT hybrid imaging increases detection rate of intra-abdominal small intestinal neuroendocrine tumor lesions. *World J Surg*. 2018;42(2):498–505.
- 21 d'Assignies G, Fina P, Bruno O, Vullierme MP, Tubach F, Paradis V, et al. High sensitivity of diffusion-weighted MR imaging for the detection of liver metastases from neuroendocrine tumors: comparison with T2-weighted and dynamic gadolinium-enhanced MR imaging. *Radiology*. 2013;268(2):390–9.
- 22 Anderson MA, Carpenter S, Thompson NW, Nostrant TT, Elta GH, Scheiman JM. Endoscopic ultrasound is highly accurate and directs management in patients with neuroendocrine tumors of the pancreas. *Am J Gastroenterol*. 2000;95(9):2271–7.
- 23 James PD, Tsolakakis AV, Zhang M, Belletrutti PJ, Mohamed R, Roberts DJ, et al. Incremental benefit of preoperative EUS for the detection of pancreatic neuroendocrine tumors: a meta-analysis. *Gastrointest Endosc*. 2015;81(4):848–56.e1.
- 24 Hellman P, Hennings J, Akerström G, Skogseid B. Endoscopic ultrasonography for evaluation of pancreatic tumours in multiple endocrine neoplasia type 1. *Br J Surg*. 2005;92(12):1508–12.
- 25 Braden B, Jenssen C, D'Onofrio M, Hocke M, Will U, Möller K, et al. B-mode and contrast-enhancement characteristics of small non-incident neuroendocrine pancreatic tumors. *Endosc Ultrasound*. 2017;6(1):49–54.
- 26 Sakamoto H, Kitano M, Suetomi Y, Maekawa K, Takeyama Y, Kudo M. Utility of contrast-enhanced endoscopic ultrasonography for diagnosis of small pancreatic carcinomas. *Ultrasound Med Biol*. 2008;34(4):525–32.
- 27 Eusebi LH, Thorburn D, Toumpanakis C, Frazzoni L, Johnson G, Vessal S, et al. Endoscopic ultrasound-guided fine-needle aspiration vs fine-needle biopsy for the diagnosis of pancreatic neuroendocrine tumors. *Endosc Int Open*. 2019;7(11):E1393–9.
- 28 Hedenström P. The best approach for sampling of pancreatic neuroendocrine tumors: EUS-FNA or EUS-FNB? *Endosc Int Open*. 2019;7(11):E1400–2.
- 29 Bauckneht M, Albano D, Annunziata S, Santo G, Guglielmo P, Frantellizzi V, et al. Somatostatin receptor PET/CT imaging for the detection and staging of pancreatic NET: a systematic review and meta-analysis. *Diagnostics*. 2020;10(8):598.
- 30 Geijer H, Breimer LH. Somatostatin receptor PET/CT in neuroendocrine tumours: update on systematic review and meta-analysis. *Eur J Nucl Med Mol Imaging*. 2013;40(11):1770–80.
- 31 Binderup T, Knigge U, Loft A, Federspiel B, Kjaer A. 18F-fluorodeoxyglucose positron emission tomography predicts survival of patients with neuroendocrine tumors. *Clin Cancer Res*. 2010;16(3):978–85.
- 32 Has Simsek D, Kuyumcu S, Turkmen C, Sanlı Y, Aykan F, Unal S, et al. Can complementary 68Ga-DOTATATE and 18F-FDG PET/CT establish the missing link between histopathology and therapeutic approach in gastroenteropancreatic neuroendocrine tumors? *J Nucl Med*. 2014;55(11):1811–7.
- 33 Yu R. Mahvash disease: 10 Years after discovery. *Pancreas*. 2018;47(5):511–5.
- 34 Jensen RT, Berna MJ, Bingham DB, Norton JA. Inherited pancreatic endocrine tumor syndromes: advances in molecular pathogenesis, diagnosis, management, and controversies. *Cancer*. 2008;113(7 Suppl 1):1807–43.
- 35 Gibril F, Chen YJ, Schrupp DS, Vortmeyer A, Zhuang Z, Lubensky IA, et al. Prospective study of thymic carcinoids in patients with multiple endocrine neoplasia type 1. *J Clin Endocrinol Metab*. 2003;88(3):1066–81.
- 36 Asgharian B, Chen YJ, Patronas NJ, Peghini PL, Reynolds JC, Vortmeyer A, et al. Meningiomas may be a component tumor of multiple endocrine neoplasia type 1. *Clin Cancer Res*. 2004;10(3):869–80.
- 37 McKeeby JL, Li X, Zhuang Z, Vortmeyer AO, Huang S, Pirner M, et al. Multiple leiomyomas of the esophagus, lung, and uterus in multiple endocrine neoplasia type 1. *Am J Pathol*. 2001;159(3):1121–7.
- 38 Asgharian B, Turner ML, Gibril F, Entsua LK, Serrano J, Jensen RT. Cutaneous tumors in patients with Multiple Endocrine Neoplasia type 1 (MEN1) and gastrinomas: prospective study of frequency and development of criteria with high sensitivity and specificity for MEN1. *J Clin Endocrinol Metab*. 2004;89(11):5328–36.
- 39 Lévy-Bohbot N, Merle C, Goudet P, Delemer B, Calender A, Jolly D, et al. Prevalence, characteristics and prognosis of MEN 1-associated glucagonomas, VIPomas, and somatostatinomas: study from the GTE (Groupe des Tumeurs Endocrines) registry. *Gastroenterol Clin Biol*. 2004;28(11):1075–81.
- 40 de Laat JM, Pieterman CRC, Weijmans M, Hermus AR, Dekkers OM, de Herder WW, et al. Low accuracy of tumor markers for diagnosing pancreatic neuroendocrine tumors in multiple endocrine neoplasia type 1 patients. *J Clin Endocrinol Metab*. 2013;98(10):4143–51.
- 41 Newey PJ, Newell-Price J. MEN1 surveillance guidelines: time to (Re)think? *J Endocr Soc*. 2022;6(2):bvac001.
- 42 Goudet P, Dalac A, Le Bras M, Cardot-Bauters C, Niccoli P, Lévy-Bohbot N, et al. MEN1 disease occurring before 21 years old: a 160-patient cohort study from the Groupe d'étude des Tumeurs Endocrines. *J Clin Endocrinol Metab*. 2015;100(4):1568–77.
- 43 Sadowski SM, Millo C, Cottle-Delisle C, Merkel R, Yang LA, Herscovitch P, et al. Results of (68)Gallium-DOTATATE PET/CT scanning in patients with multiple endocrine neoplasia type 1. *J Am Coll Surg*. 2015;221(2):509–17.
- 44 Panzuto F, Ramage J, Pritchard DM, van Velthuysen MLF, Schrader J, Begum N, et al. European Neuroendocrine Tumor Society (ENETS) 2023 guidance paper for gastroduodenal neuroendocrine tumours (NETs) G1-G3. *J Neuroendocrinol*. 2023;35(8):e13306.
- 45 Al-Salameh A, Cadiot G, Calender A, Goudet P, Chanson P. Clinical aspects of multiple endocrine neoplasia type 1. *Nat Rev Endocrinol*. 2021;17(4):207–24.
- 46 Sadowski SM, Triponez F. Management of pancreatic neuroendocrine tumors in patients with MEN 1. *Gland Surg*. 2015;4(1):63–8.
- 47 Nell S, Borel Rinkes IHM, Verkooijen HM, Bonsing BA, van Eijck CH, van Gooijck H, et al. Early and late complications after surgery for MEN1-related nonfunctioning pancreatic neuroendocrine tumors. *Ann Surg*. 2018;267(2):352–6.
- 48 Triponez F, Goudet P, Dosseh D, Cougard P, Bauters C, Murat A, et al. *Is surgery beneficial for MEN1 patients with small (< or = 2 cm), nonfunctioning pancreaticoduodenal endocrine tumor? An analysis of 65 patients from the GTE*. *World J Surg*. 2006;30(5):654–62; discussion 663–4.
- 49 Jensen RT, Niederle B, Mitry E, Ramage JK, Steinmuller T, Lewington V, et al. Gastrinoma (duodenal and pancreatic). *Neuroendocrinology*. 2006;84(3):173–82.
- 50 Gibril F, Schumann M, Pace A, Jensen RT. Multiple endocrine neoplasia type 1 and Zollinger-Ellison syndrome: a prospective study of 107 cases and comparison with 1009 cases from the literature. *Medicine*. 2004;83(1):43–83.
- 51 Brandt ML, Gagel RF, Angeli A, Bilezikian JP, Beck-Peccoz P, Bordi C, et al. Guidelines for diagnosis and therapy of MEN type 1 and type 2. *J Clin Endocrinol Metab*. 2001;86(12):5658–71.

- 52 Goudet P, Murat A, Cardot-Bauters C, Emy P, Baudin E, du Boullay Choplin H, et al. Thymic neuroendocrine tumors in multiple endocrine neoplasia type 1: a comparative study on 21 cases among a series of 761 MEN1 from the GTE (Groupe des Tumeurs Endocrines). *World J Surg.* 2009;33(6):1197–207.
- 53 Berna MJ, Annibale B, Marignani M, Luong TV, Corleto V, Pace A, et al. A prospective study of gastric carcinoids and enterochromaffin-like cell changes in multiple endocrine neoplasia type 1 and Zollinger-Ellison syndrome: identification of risk factors. *J Clin Endocrinol Metab.* 2008;93(5):1582–91.
- 54 Thomas-Marques L, Murat A, Delemer B, Penfornis A, Cardot-Bauters C, Baudin E, et al. Prospective endoscopic ultrasonographic evaluation of the frequency of non-functioning pancreaticoduodenal endocrine tumors in patients with multiple endocrine neoplasia type 1. *Am J Gastroenterol.* 2006;101(2):266–73.
- 55 Jensen RT, Cadiot G, Brandi ML, de Herder WW, Kaltsas G, Komminoth P, et al. ENETS Consensus Guidelines for the management of patients with digestive neuroendocrine neoplasms: functional pancreatic endocrine tumor syndromes. *Neuroendocrinology.* 2012;95(2):98–119.
- 56 Fendrich V, Langer P, Waldmann J, Bartsch DK, Rothmund M. Management of sporadic and multiple endocrine neoplasia type 1 gastrinomas. *Br J Surg.* 2007;94(11):1331–41.
- 57 Morrow EH, Norton JA. Surgical management of Zollinger-Ellison syndrome; state of the art. *Surg Clin North Am.* 2009;89(5):1091–103.
- 58 Fendrich V, Waldmann J, Bartsch DK, Langer P. Surgical management of pancreatic endocrine tumors. *Nat Rev Clin Oncol.* 2009;6(7):419–28.
- 59 Gibril F, Venzon DJ, Ojeaburu JV, Bashir S, Jensen RT. Prospective study of the natural history of gastrinoma in patients with MEN1: definition of an aggressive and a nonaggressive form. *J Clin Endocrinol Metab.* 2001;86(11):5282–93.
- 60 Norton JA, Fang TD, Jensen RT. Surgery for gastrinoma and insulinoma in multiple endocrine neoplasia type 1. *J Natl Compr Canc Netw.* 2006;4(2):148–53.
- 61 Hammel PR, Vilgrain V, Terris B, Penfornis A, Sauvanet A, Correas JM, et al. Pancreatic involvement in von Hippel-Lindau disease. The Groupe Francophone d'Etude de la Maladie de von Hippel-Lindau. *Gastroenterology.* 2000;119(4):1087–95.
- 62 Blansfield JA, Choyke L, Morita SY, Choyke PL, Pingpank JF, Alexander HR, et al. Clinical, genetic and radiographic analysis of 108 patients with Von Hippel-Lindau disease (VHL) manifested by pancreatic neuroendocrine neoplasms (PNETs). *Surgery.* 2007;142(6):814–8; discussion 818.e1-2.
- 63 Libutti SK, Choyke PL, Alexander HR, Glenn G, Bartlett DL, Zbar B, et al. Clinical and genetic analysis of patients with pancreatic neuroendocrine tumors associated with von Hippel-Lindau disease. *Surgery.* 2000;128(6):1022–7; discussion 1027-8.
- 64 Tirosh A, Sadowski SM, Linehan WM, Libutti SK, Patel D, Nilubol N, et al. Association of VHL genotype with pancreatic neuroendocrine tumor phenotype in patients with von Hippel-Lindau disease. *JAMA Oncol.* 2018;4(1):124–6.
- 65 Laks S, van Leeuwen R, Patel D, Keutgen XM, Hammel P, Nilubol N, et al. Management recommendations for pancreatic manifestations of von Hippel-Lindau disease. *Cancer.* 2022;128(3):435–46.
- 66 Keutgen XM, Hammel P, Choyke PL, Libutti SK, Jonasch E, Kebebew E. Evaluation and management of pancreatic lesions in patients with von Hippel-Lindau disease. *Nat Rev Clin Oncol.* 2016;13(9):537–49.
- 67 Evans DG, Howard E, Giblin C, Clancy T, Spencer H, Huson SM, et al. Birth incidence and prevalence of tumor-prone syndromes: estimates from a UK family genetic register service. *Am J Med Genet.* 2010;152a(2):327–32.
- 68 Crino PB, Nathanson KL, Henske EP. The tuberous sclerosis complex. *N Engl J Med.* 2006;355(13):1345–56.
- 69 Eledrisi MS, Stuart CA, Alshanti M. Insulinoma in a patient with tuberous sclerosis: is there an association? *Endocr Pract.* 2002;8(2):109–12.