

# Strongyloides stercoralis and immunosuppression in dermatology

## Strongyloides stercoralis e a imunossupressão na dermatologia

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### Abstract

Dermatologists can take advantage of numerous immunosuppressive drugs to treat several conditions such as autoimmune bullous dermatoses, psoriasis, and connective tissue diseases. In particular, corticosteroids often play an important role in the management of these diseases. However, prior to the start of immunosuppressive therapy, screening for opportunistic infections is crucial. Strongyloidiasis is one such disease. The parasite *Strongyloides stercoralis* is a nematode with a complex life cycle and the ability to autoinfect its host. Although it currently is a rare disease in Portugal, it has a widespread distribution especially amongst low-income countries. It is usually responsible for a chronic asymptomatic infection, albeit frequently with intermittent eosinophilia. Certain comorbidities may increase the risk for hyperinfection or disseminated disease. Such factors are the presence of immunocompromising conditions such as haematological malignancies, AIDS, HTLV-1 infection and therapies such as transplantation and corticosteroids. The screening and diagnosis are usually performed with parasitological and serological tests, and the treatment of choice is ivermectin. As such, since chronic infection can be asymptomatic and hyperinfection potentially lethal, screening prior to the start of immunosuppressive treatment is imperative. Dermatologists that prescribe such regimens should be familiar with the need of parasite screening and management prior to the start of therapy.

**Keywords:** Strongyloidiasis. *Strongyloides stercoralis*. Corticosteroid therapy. Immunosuppression in dermatology. Immunosuppression. Hyperinfection syndrome.

### Resumo

A Dermatologia tem à sua disposição inúmeros fármacos imunossupressores para tratar várias doenças como dermatoses bolhosas autoimunes, psoríase e doenças do tecido conjuntivo. Em particular, a corticoterapia tem um papel frequentemente importante na gestão destas patologias. No entanto, previamente ao início da terapêutica imunossupressora, o rastreio de infeções oportunistas, como a estromiloidíase, é crucial. O parasita *Strongyloides stercoralis* é um nematoda com um ciclo de vida complexo e com a capacidade de provocar autoinfecção no hospedeiro. Apesar de ser atualmente uma doença rara em Portugal, tem uma distribuição generalizada, sobretudo em países de baixo rendimento económico. Geralmente, é responsável por uma infeção crónica e assintomática, se bem que frequentemente com eosinofilia intermitente. Certas comorbilidades

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Received: 01-07-2022

Accepted: 28-08-2022  
DOI: 10.24875/PJDV.M22000044

Available online: 24-10-2022

Port J Dermatol and Venereol. 2022;80(3):206-212  
[www.portuguesejournalofdermatology.com](http://www.portuguesejournalofdermatology.com)

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podem aumentar o risco de hiperinfecção ou doença disseminada. Estes fatores são a presença de certas patologias com compromisso do sistema imunitário, como neoplasias hematológicas, Sida e infecção pelo HTLV-1. Também certas intervenções, como a corticoterapia ou a transplantação, são fatores de risco. O rastreio e diagnóstico fazem-se habitualmente com testes serológicos e parasitológicos, e o tratamento de escolha é a ivermectina. Assim, dado que a infecção crónica pode frequentemente ser assintomática e a hiperinfecção potencialmente letal, o rastreio prévio ao início de tratamento imunossupressor é fundamental. Neste contexto, Dermatologistas que prescrevem tais fármacos devem estar familiarizados com a necessidade de rastreio para a infecção por este parasita e sua gestão antes do início da terapêutica.

**Palavras-chave** : Estrongiloidíase. *Strongyloides stercoralis*. Terapêutica corticosteroide. Imunossupressão em dermatologia. Imunossupressão. Síndrome de hiperinfecção.

## Introduction

Dermatologists have at their disposal numerous immunosuppressive drugs that are useful in controlling several conditions such as autoimmune bullous dermatoses, psoriasis, and connective tissue disease<sup>1,2</sup>. Indeed, it is imperative that physicians are aware of the iatrogenic increased risk and severity of infection. As such, screening for several latent microorganisms can be valuable (e.g., tuberculosis, hepatitis B and C, deep fungal infections, or HIV)<sup>1,2</sup>.

One such parasite is *Strongyloides stercoralis*. It is a skin-penetrating intestinal nematode with a complex life cycle<sup>3</sup>. It is widely distributed around the world especially around the tropics<sup>3</sup> and is one of the few helminths with the ability of autoinfection<sup>4</sup>. Since it can present as a hyperinfection syndrome that occurs especially among patients with immunosuppressive conditions or therapies<sup>4-6</sup>, it is relevant to refresh its epidemiology and pathology, with a focus on the role of its screening prior to the start of immunosuppressive therapy.

## Epidemiology

Strongyloidiasis is an emerging infection with a worldwide incidence underestimated in many countries<sup>7</sup>. It has an estimated global prevalence of over 350 million people<sup>8</sup>. While it has been traditionally described among patients from tropical and subtropical countries<sup>3,9</sup>, the prevalence of infection has been increasing not only in Caribbean, Southeast Asia, Latin America, and sub-Saharan Africa, but also in southern, eastern, and central Europe<sup>6,9</sup>. While Portugal is currently considered a nonendemic country, with infection with *S. stercoralis* being found especially among immigrants from endemic countries, the prevalence of strongyloidiasis and other helminthiasis was higher during the first three quarters of the 20<sup>th</sup> century, likely due to a lack of basic sanitation conditions<sup>10</sup>. During this time, *S. stercoralis* was found especially in the regions between the Douro and Tagus rivers<sup>10</sup>, and even very

recently, a case of a Portuguese woman presenting with strongyloidiasis was reported<sup>9</sup>, even though likely infected long ago<sup>11</sup>. Moreover, in 2001, in a cohort of children from Lisbon, 0.9% were found to be infected<sup>12</sup>.

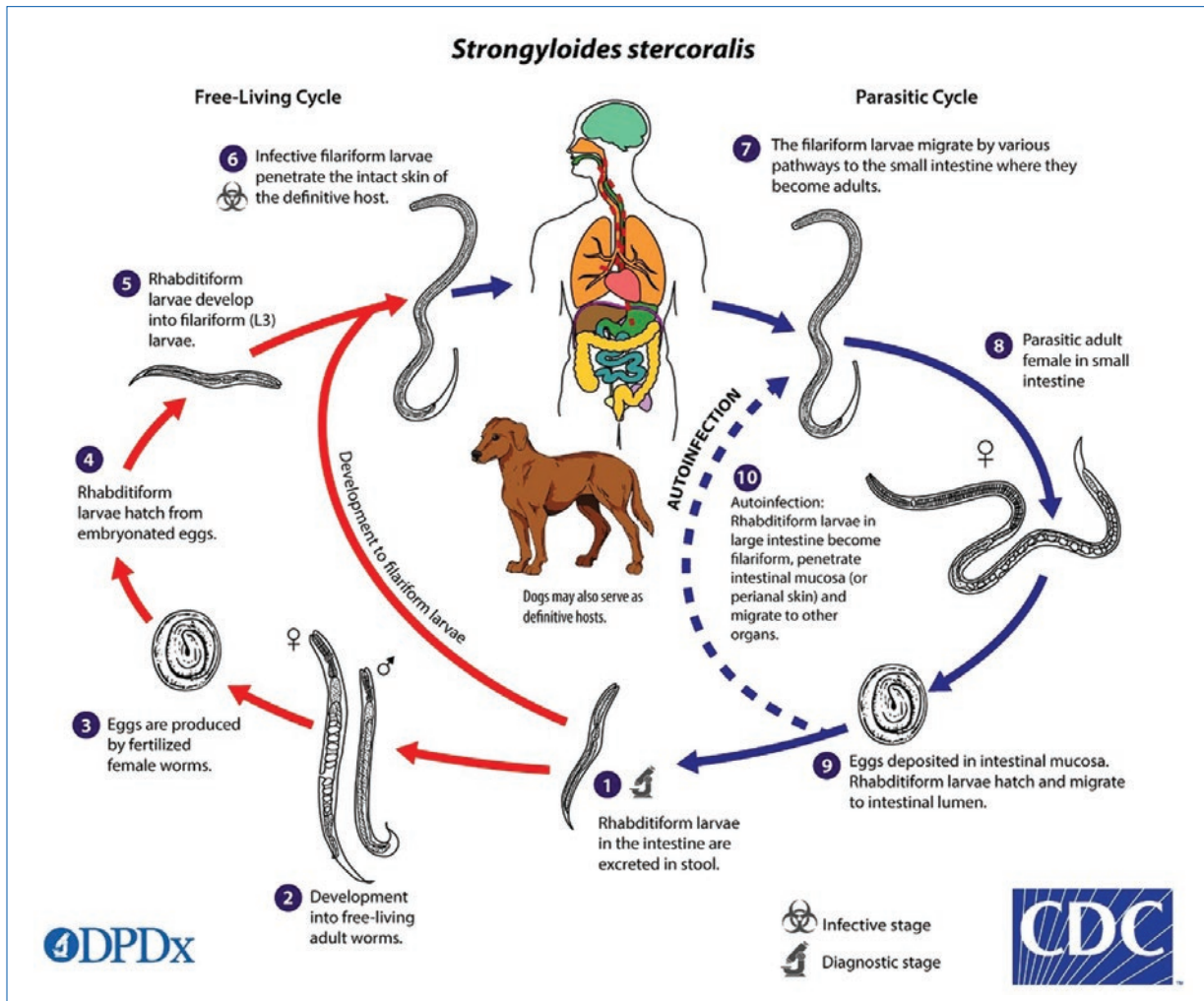
In many endemic areas, where moist soil, temperate or tropical climate and improper disposal of human waste coexist, the prevalence of strongyloidiasis can reach 50%. This is especially the case in West Africa, the Caribbean, Southeast Asia, Brazil, Cambodia, and some regions of Spain. Nevertheless, Southeast Asia seems to have the highest endemic prevalence<sup>13,14</sup>. Other risk factors for infection include white males, working with soil and travellers to areas of endemicity<sup>6</sup>. Although strongyloidiasis occurs in all ages, infection usually happens in childhood, since children are more likely to play outdoors with higher exposure to contaminated soil<sup>12,15</sup>.

Patients with certain immunosuppressive conditions are also at a higher risk for strongyloidiasis. Indeed, an altered cellular immunity (especially those on long-term corticosteroid therapy, but also human immunodeficiency virus [HIV] infection/acquired immunodeficiency syndrome), certain hematological malignancies and therapies (such as those for lymphoma and allograft transplant recipients) are at a higher risk for severe strongyloidiasis infection<sup>5,16,17</sup>. Human T-lymphotropic virus type 1 (HTLV-1) infection is also related to *S. stercoralis* with increased prevalence of this parasite in overlapping endemicity areas<sup>3,18</sup>. Indeed, corticosteroid treatment and HTLV-1 infection are the two conditions most associated with hyperinfection<sup>16</sup>.

## Lifecycle and transmission

*Strongyloides stercoralis* has a complex life cycle with two unique and distinct cycles (Fig. 1). While transmission usually occurs through contact with contaminated soil, person-to-person transmission has been described, especially among men who have sex with men<sup>19</sup>.

As one of the few helminths that is able to autoinfect its human host<sup>20</sup>, rhabditiform larvae can fertilize into



**Figure 1.** *Strongyloides stercoralis* life cycle (Image from courtesy of DPDx, a website by the Centers for Disease Control and Prevention (CDC)'s Division of Global Health, Parasitic Disease and Malaria)<sup>54</sup>. It consists of a free-living cycle in the soil, where both males and females coexist and maintain infestation in the ground. Here, eggs are hatched as rhabditiform larvae and afterward transformed into infective filariform larvae. In this stage, the larvae penetrate the skin and migrate to the small intestine where they mature into adult females and produce eggs parthenogenetically. These hatch into rhabditiform larvae that are excreted in the stool and can lead to autoinfection. These parasitic females may live up to five years, continuing their reproductive cycle<sup>6,18,20</sup>.

its filariform stage in the large bowel. Afterwards they migrate through the lymphatic and venous circulation, reaching the pulmonary circulation, alveolar space, and crawling up the respiratory tract. Then they return to the intestine through swallowed sputum<sup>6,18</sup>. External auto-infection can also occur, in which case it often leads to the development of larva currens<sup>6</sup>.

Almost all strongyloidiasis are due to infection with *Strongyloides stercoralis*. However, the primate parasite *Strongyloides fülleborni* has been described in children in Africa and in Papua New Guinea, where it is a cause of “swollen belly syndrome”<sup>20</sup>.

### Pathogenesis and clinical manifestations

*Strongyloides stercoralis* infection was first described in 1876 from the stool of French soldiers with diarrhea who were returning from the old Indochina region, leading to the designation of “Cochin-China diarrhoea”<sup>21</sup>. Manifestations of primary acute infection with *Strongyloides stercoralis* are directly related to its life cycle. After skin penetration, if the larvae do not find their natural route to the circulation and stay in the integument, larva migrans presenting as a maculopapular, pruriginous and serpiginous rash can occur<sup>22</sup>.

**Table 1.** Principal manifestations of acute and chronic strongyloidiasis<sup>7,9,20,22,23,26,55</sup>. Strongyloidiasis presentation directly relates to the parasite's life cycle. Acute infection can be asymptomatic in up to one-third of infections. Chronic strongyloidiasis is often asymptomatic, with eosinophilia being the sole, albeit intermittent, marker. Otherwise, gastrointestinal symptoms can occur, with larva currens, abdominal pain, and diarrhea being a classically recognized triad

	Acute strongyloidiasis	Chronic strongyloidiasis
Gastrointestinal manifestations	<ul style="list-style-type: none"> <li>• Abdominal pain, malabsorption, steatorrhea, diarrhea</li> <li>• Onset usually 2 weeks after infection; larvae found on the stool after 3-4 weeks</li> </ul>	<ul style="list-style-type: none"> <li>• Diarrhea, malabsorption, steatorrhea, constipation, abdominal pain, intermittent vomiting</li> </ul>
Respiratory manifestations	<ul style="list-style-type: none"> <li>• Cough, wheezing, shortness of breath, tracheal irritation, bronchitis, Loeffler's syndrome, transient pulmonary infiltrates</li> <li>• Onset a few days after infection</li> </ul>	<ul style="list-style-type: none"> <li>• Cough, dyspnoea, recurrent asthma</li> <li>• Often mild or absent</li> </ul>
Other	<ul style="list-style-type: none"> <li>• Larva migrans, fever, anorexia</li> <li>• Eosinophilia (as high as 75-80%)</li> </ul>	<ul style="list-style-type: none"> <li>• Intermittent eosinophilia and elevated IgE levels, often isolated</li> <li>• Nephrotic syndrome</li> <li>• Pruritus ani, larva currens, urticarial, petechial and purpuric rashes</li> </ul>

Manifestations of acute and chronic strongyloidiasis can be found in Table 1. Chronic infection is often asymptomatic, with eosinophilia being the sole, albeit intermittent, marker<sup>9,23</sup>. Actually, hypersensitivity is an important part of the immune response to this parasite, contributing both to the pathogenesis of the disease and to its protection<sup>16</sup>. In fact, a primary Th2 response favors infection by increasing tissue permeability to the parasite and reducing complement activation, important for the larvae-killing capabilities of eosinophils and granulocytes<sup>24,25</sup>, but interleukin-13 causes increased peristalsis, possibly leading to increased larval expulsion<sup>25</sup>. On the other hand, HTLV-1 infection, a known risk factor for severe strongyloidiasis, results in an increased interferon-gamma production and decreased levels of interleukin-4 and IgE, which creates a favorable environment for *Strongyloides stercoralis* proliferation<sup>16</sup>.

Whereas internal autoinfection is usually less relevant in healthy individuals, in immunosuppressed patients it can present as one of the two most severe forms of strongyloidiasis, either the hyperinfection syndrome or the disseminated disease. Although immunocompetent patients are also at risk, those with impaired cell-mediated immunity are much more susceptible<sup>6,26</sup>. In severe strongyloidiasis in the immunocompromised host, eosinophilia is often absent<sup>23</sup>. In the hyperinfection syndrome there is a favorable environment for parasitic proliferation, resulting in an increased burden along the usual migration pattern. It essentially is an accelerated auto-infection and the distinction between these two is merely quantitative and not strictly defined<sup>26</sup>.

As such, new onset or exacerbation of gastrointestinal and pulmonary symptoms is frequent, and the identification of increased numbers of larvae in faeces and/or

respiratory samples is the hallmark of hyperinfection<sup>26</sup>. While the increased numbers can lead to complications such as intestinal obstruction, ileus, and gastrointestinal bleeding, usually there is no metastatic dissemination outside the regular migration pattern. Nevertheless, migration of larvae that carry bacteria on the surface of the larval integument, as excreta from the larval intestinal tract<sup>27</sup> or the presence of ulcers may facilitate the spread and systemic infection with enteric bacteria<sup>26</sup>.

Pulmonary complications including pulmonary infiltrates, diffuse alveolar hemorrhage, and respiratory failure can develop in patients with hyperinfection syndrome and, if not treated, may be lethal. Indeed, a lack of familiarity with this parasite leading to delayed screening and treatment is a cause for a high mortality among immunosuppressed patients<sup>28</sup>.

While hyperinfection denotes an increased parasite replication, disseminated strongyloidiasis implies widespread dissemination to extraintestinal organs, without the obligatory need for an increased parasite proliferation or severity of disease<sup>26</sup>. Multiple organs beyond the range of its normal life cycle are affected, including the liver, heart, kidneys, and central nervous system<sup>6</sup>. In severe disease, and as in hyperinfection, translocation of enteric bacteria can occur, leading to polymicrobial bacteremia or meningitis<sup>6</sup>. Cerebrospinal fluid analysis shows neutrophilic pleocytosis with an elevated protein level and low glucose level. A gram stain can be positive for enteric bacteria and direct examination can reveal *Strongyloides stercoralis* larvae<sup>29,30</sup>.

Other manifestations include lymphadenopathy, fever, haemoptysis, cough, anaemia, vomiting, weight loss, abdominal pain, and distension<sup>31</sup>. Since it coexists frequently with hyperinfection syndrome in the immunosuppressed patient, its manifestations may overlap<sup>6</sup>.

## Diagnosis

Since most patients with strongyloidiasis do not present with distinct clinical features, the diagnosis requires a high degree of suspicion<sup>11</sup>.

*Strongyloides stercoralis* larvae can be intermittently found in faeces usually a month after skin penetration. Usually, only larvae are found since the eggs immediately hatch in the intestine. *Strongyloides fulleborni*, however, sheds eggs in faeces, and is readily found using microscopy<sup>6</sup>. Direct smear examination of stool in saline and Lugol's iodine stain is a definitive diagnostic testing, although with a low sensitivity (as low as 21%). However, concentration methods, such as formalin-ethyl acetate, Harada-Mori techniques, and Baermann concentration increase the yield and are significantly more sensitive<sup>32,33</sup>. While diagnosis of hyperinfection is relatively easy due to the high quantity of larvae in stool and sputum, outside of this setting it is often inadequate, as a single stool examination is less than 50% sensitive for making diagnosis<sup>34</sup>. As such, it is mandatory to screen multiple times, ideally using a concentration method, although they are seldom performed in most parasitology labs<sup>34,35</sup>. A sensitivity higher than 90% can be achieved if seven or more samples are examined<sup>35</sup>. When concerning the hyperinfection syndrome, the examination of a duodenal aspirate for eggs and larva is the most sensitive diagnostic procedure (as high as 90%)<sup>31</sup>.

In addition to faeces samples, endoscopic examination and biopsies can be useful. Endoscopy may range from normal-appearing mucosa to severe duodenitis or colitis with oedematous and erythematous mucosa and white villi. Moreover, in hyperinfection with pulmonary involvement, larvae can be shown in duodenal biopsy<sup>36</sup>. In disseminated disease, larvae can be found in several extraintestinal sites, such as skin biopsy, cerebrospinal fluid, urine, pericardial, pleural and peritoneal fluid<sup>37–42</sup>.

Serological assays are another useful tool in the diagnosis and follow-up of strongyloidiasis. Specific antibodies can be used as a follow-up to prove seroconversion after a successful therapy. There are several commercially available tests with varying sensitivities and specificities. For example, ELISA seems to be a sensitive test (88–95%), albeit with a variable specificity (29–99%)<sup>34</sup>. The low specificity is due to cross-reactivity with other helminth infections, such as filariasis, ascariasis and acute schistosomiasis<sup>34</sup>. However, in Portugal, these are likely not frequent differential diagnosis, and this appear to be a smaller issue with more recent test kits<sup>43</sup>. Another drawback of

these tests is their lower sensitivity in severely immunosuppressed patients, and incapacity to accurately distinguish between past and present infection among patients already treated for strongyloidiasis or originating from an endemic country<sup>44</sup>. However, antibody titres tend to diminish with time, although the time required to become negative may be higher than 12 months<sup>5</sup>.

Real-time polymerase chain reaction is another tool for the diagnosis of strongyloidiasis, albeit not being readily available in most centres. Estimates of sensitivity of this method are variable but seem high. In the future, molecular testing may enhance the diagnosis of this infection<sup>6,34</sup>.

## Screening and management

Strongyloidiasis should be a differential diagnosis in any patient with unexplained eosinophilia, especially if there was exposure in endemic areas. However, immunocompetent patients with high risk of exposure should still be screened, even if without eosinophilia<sup>5</sup>. Moreover, in patients with risk factors for developing hyperinfection, testing should also be considered, particularly when having a history of originating or travelling to an endemic country, even if in a distant past<sup>5,11,25</sup>. This is especially important in patients that have immunosuppressive conditions or treatments, such as those with hematologic malignancies, undergoing transplantation or corticosteroid therapy<sup>5</sup>. Indeed, in this case, both parasitological and serological assays should be used<sup>5,25</sup>. In some cases, pre-emptive ivermectin treatment should be considered, if a diagnostic test is not available<sup>5</sup>. However, although corticosteroid exposure has been identified as the main risk factor, there are also reports regarding the use of non-steroid immunosuppressive agents and biologic therapies, including those directed at IL-1, TNF $\alpha$  and lymphocyte depleting drugs<sup>45,46</sup>. Nevertheless, while IgE, IL-13, and IL-4 are paramount for the pathogenesis of this disease, unexpectedly, the modulation of these cytokines has not yet been found to increase risk of strongyloidiasis. Still, pre-treatment screening is advised<sup>47,48</sup>. Additionally, screening should also be considered in those with evidence of HTLV-1 infection<sup>49</sup>.

Dermatologists can take advantage of numerous immunosuppressive drugs in the management of several ailments such as chronic immunoinflammatory diseases, psoriasis, and connective tissue disease. It is, therefore, imperative that a screening for the relevant opportunistic diseases be considered prior to the start of treatment. Strongyloidiasis is one such illness, and

several guidelines recommend its screening prior to the start of several medications, primarily corticosteroids<sup>1,2,50</sup>. To do so, likely a combination of serological and microbiological assays is ideal, since in general, serology is highly sensitive, while stool examination is highly specific. Moreover, immunosuppressed patients may have a lower serological sensitivity which might be overcome by an increased detection in stool samples<sup>45,51</sup>. As such, a pre-treatment screen with several (perhaps more than seven) stool samples and serology is advised.

Treatment of strongyloidiasis is usually performed with ivermectin. This broad-spectrum antiparasitic causes muscle paralysis in invertebrates by activating chlorine channels<sup>52</sup>. It is better tolerated and has a similar efficacy than thiabendazole, and is more effective than albendazole<sup>5,25</sup>. In uncomplicated infections a single 200 µg/kg/day oral dose of ivermectin for one or two days usually sufficient<sup>5</sup>. A repetition of this course could be suggested after two weeks, to account for the parasite's autoinfective cycle, however a randomized clinical trial failed to show advantage in this strategy<sup>53</sup>.

When considering hyperinfection, there is a lack in high-quality evidence. However, it has been suggested that ivermectin should be given daily or every 48 hours at a dose of 200 µg/kg/day for at least one to two weeks. When the oral route is not well tolerated, alternative routes can be considered. Multiple follow-up stool assessments should be performed, and treatment continuation until no more larvae are found in faeces should be considered<sup>7,25</sup>. Additionally, whenever iatrogenic immunosuppression is present, reduction in these regimens should be considered, when clinically feasible<sup>25</sup>. Moreover, these patients should be considered infectious and put under standard contact precautions<sup>49</sup>.

The high mortality in hyperinfection is often due to a lack of awareness in the need for parasite screening before the start of corticosteroid therapy<sup>28</sup>. Indeed, several patients with fatal outcomes after treatment with empirical corticosteroids are later confirmed as a case of disseminated strongyloidiasis. Moreover, the possibility of infection with this nematode should be considered in any immunocompromised patient who suddenly deteriorates without any apparent cause, since delay in treatment often results in death<sup>14</sup>.

## Conclusion

Currently, strongyloidiasis is a rare disease in Portugal, mostly related to migrant population. However, it can have a severe if not fatal course, especially

amongst immunosuppressed patients. As such, and since chronic infection can often be asymptomatic, screening prior to the start of immunosuppressive treatment (especially corticosteroids) is imperative. Dermatologists that prescribe such regimens should be familiar with the need of parasite screening and management prior to the start of therapy.

## Ethical disclosures

**Protection of human and animal subjects.** The authors declare that no experiments were performed on humans or animals for this study.

**Confidentiality of data.** The authors declare that no patient data appear in this article.

**Right to privacy and informed consent.** The authors declare that no patient data appear in this article.

## References

1. Lehman JS, Wetter DA, Davis MDP, El-Azhary RA, Gibson LE, Kalaaji AN. Anticipating and preventing infection in patients treated with immunosuppressive medications for dermatologic indications: a dermatologist's checklist. *J Am Acad Dermatol* 2014;71:e125–6. DOI: 10.1016/j.jaad.2014.03.021
2. Ponzo MG, Hong CH. A dermatologist's guide to infection screening prior to initiating immunosuppressive therapy. *Skin Therapy Lett* 2017;22:8–11.
3. Schär F, Trostorf U, Giardina F, Khieu V, Muth S, Marti H, et al. Strongyloides stercoralis: global distribution and risk factors. *PLoS Negl Trop Dis* 2013;7:e2288.
4. Valerio L, Roure S, Fernández-Rivas G, Basile L, Martínez-Cuevas O, Ballesteros AL, et al. Strongyloides stercoralis, the hidden worm. Epidemiological and clinical characteristics of 70 cases diagnosed in the North Metropolitan Area of Barcelona, Spain, 2003–2012. *Trans R Soc Trop Med Hyg* 2013;107:465–70. DOI: 10.1093/trstmh/trt053
5. Requena-Méndez A, Buonfrate D, Gomez-Junyent J, Buonfrate D, Zammarchi L, Requena-Méndez A, et al. Evidence-based guidelines for screening and management of strongyloidiasis in non-endemic countries. *Am J Trop Med Hyg* 2017;97:645–52. DOI: 10.4269/ajtmh.16-0923
6. Puthiyakunnon S, Boddu S, Li Y, Zhou X, Wang C, Li J, et al. Strongyloidiasis—an insight into its global prevalence and management. Simon GL, editor *PLoS Negl Trop Dis* 2014;8:e3018. DOI: 10.1371/journal.pntd.0003018
7. Montes M, Sawhney C, Barros N. Strongyloides stercoralis: there but not seen. *Curr Opin Infect Dis* 2010;23:500–4. DOI: 10.1097/QCO.0b013e32833df718
8. Bisoffi Z, Buonfrate D, Montresor A, Requena-Méndez A, Muñoz J, Krolewiecki AJ, et al. Strongyloides stercoralis: a plea for action. *PLoS Negl Trop Dis* 2013;7:e2214. DOI: 10.1371/journal.pntd.0002214
9. Pinto J, Almeida P, Meireles D, Araújo A. Strongyloidiasis: a diagnosis to consider in previously endemic regions in Portugal. *Acta Med Port* 2021;34:552–6. DOI: 10.20344/amp.12960
10. David de Morais J. Occurrence of native strongyloidiasis in Portugal - retrospective revision. *Rev Port Doenças Infecciosas* 2012;8:85–93.
11. Pirisi M, Salvador E, Bisoffi Z, Gobbo M, Smirne C, Gigli C, et al. Unsuspected strongyloidiasis in hospitalised elderly patients with and without eosinophilia. *Clin Microbiol Infect* 2006;12:787–92. DOI: 10.1111/j.1469-0691.2006.01500.x
12. Fernandes S, Beorlegui M, Brito MJ, Rocha G. Protocolo de parasitoses intestinais. *Acta Pediátrica Port* 2008;43:35–41.
13. Glinz D, Silué KD, Knopp S, Lohourignon LK, Yao KP, Steinmann P, et al. Comparing diagnostic accuracy of Kato-Katz, Koga Agar Plate, Ether-Concentration, and FLOTAC for Schistosoma mansoni and Soil-transmitted helminths. Geiger SM, editor *PLoS Negl Trop Dis* 2010;4:e754. DOI: 10.1371/journal.pntd.0000754
14. Johnston FH, Morris PS, Speare R, McCarthy J, Currie B, Ewald D, et al. Strongyloidiasis: a review of the evidence for Australian practitioners. *Aust J Rural Health* 2005;13:247–54. DOI: 10.1111/j.1440-1584.2005.00710.x

15. Moon TD, Oberhelman RA. Antiparasitic Therapy in Children. *Pediatr Clin North Am* 2005;52:917–48. DOI: 10.1016/j.pcl.2005.02.012
16. Evering T, Weiss LM. The immunology of parasite infections in immunocompromised hosts. *Parasite Immunol* 2006;28:549–65.
17. Ferreira MS. Strongyloidiasis and acquired immunodeficiency syndrome. In: *Enfermedades Emergentes*. 2003. p. 18–26.
18. Ye L, Taylor GP, Rosadas C. Human T-cell lymphotropic virus type 1 and *Strongyloides stercoralis* co-infection: a systematic review and meta-analysis. *Front Med* 2022;9:1–9. DOI: 10.3389/fmed.2022.832430
19. Sorvillo F, Mori K, Sewake W, Fishman L. Sexual transmission of *Strongyloides stercoralis* among homosexual men. *Br J Vener Dis* 1983;59:342. DOI: 10.1136/sti.59.5.342
20. Clark T, Gilman R. Hookworm and *Strongyloides* Infections. In: Ryan E, Hill D, Solomon T, Aronson N, Endy T, editors. *Hunter's Tropical Medicine and Emerging Infectious Diseases*. 10th ed. Elsevier Inc; 2020. p. 845–50.
21. Normand LA. Sur la maladie dite diarrhée de Cochinchine. *Comptes Rendus l'Académie des Sci* 1983;83:316.
22. Corte LD, da Silva MVS, Souza PRM. Simultaneous larva migrans and larva currens caused by *Strongyloides stercoralis*: a case report. *Case Rep Dermatol Med* 2013;2013:1–3. DOI: 10.1155/2013/381583
23. Grove D. Clinical Manifestations. In: Grove D, editor. *Strongyloidiasis: a major roundworm infection of man*. Taylor & Francis; 1989. p. 155–73.
24. Bakiri AH, Mingomataj EC. Parasites induced skin allergy: a strategic manipulation of the host immunity. *J Clin Med Res* 2010;2:247–55. DOI: 10.4021/jocmr456w
25. Czeresnia JM, Weiss LM. *Strongyloides stercoralis*. *Lung* 2022;200:141–8. DOI: 10.1007/s00408-022-00528-z
26. Keiser PB, Nutman TB. *Strongyloides stercoralis* in the immunocompromised population. *Clin Microbiol Rev* 2004;17:208–17. DOI: 10.1128/CMR.17.1.208-217.2004
27. Scowden EB, Schaffner W, Stone WJ. Overwhelming strongyloidiasis: an unappreciated opportunistic infection. *Medicine (Baltimore)* 1978;57:527–44.
28. Boulware DR, Stauffer WM, Hendel-Paterson BR, Rocha JLL, Seet RCS, Summer AP, et al. Maltreatment of *Strongyloides* infection: case series and worldwide physicians-in-training survey. *Am J Med* 2007;120:545.e1–8. DOI: 10.1016/j.amjmed.2006.05.072
29. Celedon JC, Mathur-Wagh U, Fox J, Garcia R, Wiest PM. Systemic strongyloidiasis in patients infected with the human immunodeficiency virus: A report of 3 cases and review of the literature. *Med (United States)* 1994;73:256–63. DOI: 10.1097/00005792-199409000-00004
30. Jain AK, Agarwal SK, El-Sadr W. *Streptococcus bovis* bacteremia and meningitis associated with *Strongyloides stercoralis* colitis in a patient infected with human immunodeficiency virus. *Clin Infect Dis* 1994;18:253–4. DOI: 10.1093/clinids/18.2.253
31. Ramdial PK, Hlatshwayo NH, Singh B. *Strongyloides stercoralis* mesenteric lymphadenopathy: clue to the etiopathogenesis of intestinal pseudo-obstruction in HIV-infected patients. *Ann Diagn Pathol* 2006;10:209–14. DOI: 10.1016/j.anndiagpath.2005.11.008
32. Kemp L, Hawley T. Strongyloidiasis in a hyperinfected patient. *Lab Med* 1996;27:237–40.
33. Campo Polanco L, Gutiérrez LA, Cardona Arias J. Infección por *Strongyloides stercoralis*: metanálisis sobre evaluación de métodos diagnósticos convencionales (1980-2013). *Rev Esp Salud Publica* 2014;88:581–600.
34. Siddiqui AA, Berk SL. Diagnosis of *Strongyloides stercoralis* infection. *Clin Infect Dis* 2001;33:1040–7. DOI: 10.1086/322707
35. Nielsen PB, Mojon M. Improved diagnosis of strongyloides stercoralis by seven consecutive stool specimens. *Zentralbl Bakteriol Mikrobiol Hyg A* 1987;263:616–8. DOI: 10.1016/s0176-6724(87)80207-9
36. Kishimoto K. Endoscopic and histopathological study on the duodenum of *Strongyloides stercoralis* hyperinfection. *World J Gastroenterol* 2008;14:1768.
37. Coovadia YM, Rajput MC, Bhana RH. Disseminated strongyloidiasis in a diabetic patient. *Trop Geogr Med* 1993;45:179–80.
38. Premanand R, Prasad GV, Mohan A, Gururajkumar A, Reddy MK. Eosinophilic pleural effusion and presence of filariform larva of *Strongyloides stercoralis* in a patient with metastatic squamous cell carcinoma deposits in the pleura. *Indian J Chest Dis Allied Sci* 2003;45:121–4.
39. Murali A, Rajendiran G, Ranganathan K, Shanthakumari S. Disseminated infection with *Strongyloides stercoralis* in a diabetic patient. *Indian J Med Microbiol* 2010;28:407–8. DOI: 10.3748/wjg.14.1768
40. Sebastian IA, Pandian JD, Oberoi A, Kate M, Jaison V, Bose S, et al. Disseminated Strongyloidiasis: breaking brain barriers. *Ann Indian Acad Neurol* 2019;22:234–7. DOI: 10.4103/aiian.AIAN\_321\_18
41. Barreto NMPV, De Souza JN, Araújo WAC, Khouri NA, De Oliveira EP, Teixeira MCA, et al. Urinary tract infection by *Strongyloides stercoralis*: a case report *J Parasitol* 2018;104:433–7. DOI: 10.1645/17-88
42. Gordon SM, Gal AA, Solomon AR, Bryan JA. Disseminated strongyloidiasis with cutaneous manifestations in an immunocompromised host. *J Am Acad Dermatol* 1994;31:255–9. DOI: 10.1016/s0190-9622(94)70158-x
43. Requena-Méndez A, Chiodini P, Bisoffi Z, Buonfrate D, Gotuzzo E, Muñoz J. The laboratory diagnosis and follow up of strongyloidiasis: a systematic review. *PLoS Negl Trop Dis* 2013;7:e2002. DOI: 10.1371/journal.pntd.0002002</>
44. Gam AA, Neva FA, Krotoski WA. Comparative sensitivity and specificity of ELISA and IHA for serodiagnosis of strongyloidiasis with larval antigens. *Am J Trop Med Hyg* 1987;37:157–61. DOI: 10.4269/ajtmh.1987.37.157
45. Boggild AK, Libman M, Greenaway C, McCarthy A. CATMAT statement on disseminated strongyloidiasis: prevention, assessment and management guidelines. *Can Commun Dis Rep* 2016;42:12–9. DOI: 10.14745/ccdr.v42i01a03
46. Gétaz L, Castro R, Zamora P, Kramer M, Gareca N, Torrico-Espinoza M del C, et al. Epidemiology of *Strongyloides stercoralis* infection in Bolivian patients at high risk of complications *PLoS Negl Trop Dis* 2019;13:e0007028. DOI: 10.1371/journal.pntd.0007028
47. Braddock M, Hanania NA, Sharafkhaneh A, Colice G, Carlsson M. Potential Risks related to modulating interleukin-13 and interleukin-4 signalling: a systematic review. *Drug Saf* 2018;41:489–509. DOI: 10.1007/s40264-017-0636-9
48. Winthrop KL, Mariette X, Silva JT, Benamu E, Calabrese LH, Dumusc A, et al. ESCMID Study Group for Infections in Compromised Hosts (ESGICH) Consensus Document on the safety of targeted and biological therapies: an infectious diseases perspective (Soluble immune effector molecules [I]): agents targeting interleukins, immunoglobulins. *Clin Microbiol Infect* 2018;24:S21–40. DOI: 10.1016/j.cmi.2018.02.002
49. Mejia R, Nutman TB. Screening, prevention, and treatment for hyperinfection syndrome and disseminated infections caused by *Strongyloides stercoralis*. *Curr Opin Infect Dis* 2012;25:458–63. DOI: 10.1097/QCO.0b013e3283551dbd
50. Murrell DF. Screening for infections prior to initiating immunosuppressive treatment for patients with autoimmune blistering diseases. *Br J Dermatol* 2014;171:1285–6. DOI: 10.1111/bjd.13451
51. Carnino L, Schwob JM, Gétaz L, Nickel B, Neumayr A, Eperon G. A practical approach to screening for *Strongyloides stercoralis*. *Trop Med Infect Dis* 2021;6:203. DOI: 10.3390/tropicalmed6040203
52. McCarthy J, Loukas A, Hotez P. Chemotherapy of Helminth Infections. In: Brunton L, Chabner B, Knollman B, editors. *Goodman & Gilman's The Pharmacological Basis of Therapeutics*. 12th ed. McGraw-Hill Medical; 2011.
53. Greaves D, Coggle S, Pollard C, Aliyu SH, Moore EM. *Strongyloides stercoralis* infection. *BMJ* 2013;347:f4610. DOI:10.1136/bmj.f4610
54. Centers for Disease Control and Prevention *Strongyloidiasis* [Internet] 2019 [cited 2022 Jun 6] Available from: <https://www.cdc.gov/dpdx/strongyloidiasis/index.html>
55. Al Hadidi M, Shaaban H, Jumean K, Peralta P. Loeffler's Syndrome secondary to hyperinfection by *Strongyloides stercoralis* Associated with methotrexate in a patient with rheumatoid arthritis. *J Glob Infect Dis* 2018;10:29–30. DOI: 10.4103/jgid.jgid\_69\_17