

Lavandula stoechas subsp. *luisieri* and *L. pedunculata* - phytochemical study, micromorphology and histochemistry

Lavandula stoechas subsp. *luisieri* e *L. pedunculata* - estudo fitoquímico, micromorfologia e histoquímica

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ABSTRACT

Extracts of different polarities were obtained from plants of *Lavandula stoechas* subsp. *luisieri* and *Lavandula pedunculata* collected in the Center and Southwest Portugal. In order to have an idea on the possible classes of compounds present in each extract, they were subjected to a preliminary phytochemical screening through thin layer chromatography on silica gel plates, developed in appropriated mixtures of eluents and revealed with the specific reagents. Both *taxa* showed a hydrophilic and lipophilic chemical composition, being more diverse in *L. stoechas* subsp. *luisieri*. The micromorphology and histochemistry of secretory structures were also investigated by light and scanning electron microscopy. The common types of glandular trichomes, found in the Lamiaceae, peltate and capitate type I and II, were identified in each species as well as non-glandular stellate trichomes. In addition to these, *L. pedunculata* also presented capitate type III trichomes and still more two types of secretory trichomes: i) stellate type with both glandular and non glandular arms and ii) another type of capitate trichome, with a long and broad stalk formed by 2 cells, the basal is conical and much larger than the second, this similar to a ring, which supports a pluricellular head. These types of trichomes appear in the leaves but predominate in the flowers. The histochemical tests confirmed the existence of hydrophilic and lipophilic metabolites in glandular trichomes, except in the peltate hairs of the leaves of *L. stoechas* subsp. *luisieri* where only lipophilic secretions prevail.

Keywords: *Lavandula*, trichomes, phytochemical study, micromorphology, histochemistry

RESUMO

Extratos de diferentes polaridades foram obtidos de plantas de *Lavandula stoechas* subsp. *luisieri* e *L. pedunculata*, colhidas no centro e sudoeste de Portugal. Para ter uma ideia sobre as possíveis classes de compostos presentes em cada extracto, estes foram submetidos a um estudo fitoquímico preliminar através de cromatografia em camada fina, em placas de sílica gel, desenvolvidos em misturas apropriadas de eluentes e reveladas com reagentes específicos. Os dois *taxa* apresentaram uma composição química de natureza hidrofílica e lipofílica, sendo mais diversificada e encontrada em *Lavandula stoechas* subsp. *luisieri*. A micromorfologia e histoquímica das estruturas secretoras foram observadas através de microscopia ótica e eletrónica de varrimento. Os tipos comuns de tricomas glandulares encontrados nas Lamiaceae, tricomas peltados e capitados tipo I e II, foram identificados nos 2 *taxa*, assim como tricomas não-glandulares de tipo estrelado. Além destes, *L. pedunculata* apresentou ainda tricomas capitados tipo III e mais dois tipos de tricomas secretores: i) estrelado com braços glandulares ou estrelado com braços não glandulares e glandulares e ii) um outro tipo de tricoma capitado, de pedúnculo longo e largo, formado por 2 células, a basal cónica, muito maior do que a 2ª célula, esta semelhante a um anel, que suporta uma cabeça pluricelular. Estes dois tipos de tricomas aparecem em folhas mas predominam nas flores. Testes histoquímicos confirmaram a existência de metabolitos secundários de natureza hidrofílica e lipofílica nos tricomas glandulares, salvo nos tricomas peltados das folhas de *L. luisieri* subsp. *stoechas*, onde predominaram apenas secreções lipofílicas.

Palavras chave: *Lavandula*, tricomas, estudo fitoquímico, micromorfologia, histoquímica

Introduction

The Lamiaceae is one of the most important botanical families found in the area of Mediterranean influence. Due to medicinal properties and economic potential in the food and fragrance industries, there is a great interest in their plant species biological and chemical characterization. Nevertheless, only *Lavandula* species of considerable economic significance are usually well documented (Cavanagh, 2002). In contrast, the native *taxa* have received much less attention and are only reported as aromatic species and little information is available on their glandular structures, their chemicals and their activities (Chizzola, 2006; Combrinck *et al.*, 2007). This is the case of some *Lavandula* species and subspecies from Portugal, among them *Lavandula stoechas* subsp. *luisieri* (Rozeira) Rozeira (Syn: *L. luisieri*; *L. stoechas* var. *luisieri*) and *Lavandula pedunculata* (Mill.) Cav. (Syn: *L. pedunculata* subsp. *sampaiana*; *L. stoechas* subsp. *pedunculata*) (Morales, 2010).

Some morphological characters and the chemical composition are known to be affected by environmental conditions and by the phenological state of the plants. These are two approaches for plant description and classification also applied to the Lamiaceae where polymorphisms and chemotypes are known (Husain *et al.*, 1990; Harley *et al.* 2004; Salmaki *et al.*, 2009). The great majority of the members of this family show non glandular and glandular trichomes, located in the epidermis of aerial parts which may vary in their density and morphology. According to the structure and mode of secretion, glandular trichomes can be classified into two general types, peltate and capitate (Werker *et al.*, 1985). The last are morphologically more diverse and can be divided into other types, also according to its structure and also to its method of secretion (Werker *et al.*, 1985).

In order to achieve a better knowledge about the chemical composition and micromorphology of the Portuguese *taxa* previously mentioned, we performed a series of increasing polarity extracts and

carried out their preliminary phytochemical screening by chromatographic methods. Light and scanning electron microscopy were used to elucidate about the secretory structures and histochemical tests were done to achieve cell and glandular localization of the main chemical groups.

Material and Methods

Plant material

Small branches of both species at flowering stage were collected between 2008 and 2011, from plants growing in open field, in the Center and Southwestern regions of Portugal. Vouchers were deposited at the Lisbon Botanical Garden Herbarium (LISU) and the Lisbon Agronomic Institute Herbarium (LISI), (Table 1). For obtaining plant extracts we used leaves and flowers of only 2 *taxa* (Table 1). For micromorphological and histochemical studies full-expanded leaves and flowers were considered.

Preparation of crude plants extracts

Crude plants extracts were prepared by extracting sequentially 100 g of dried powdered plant material with 300 ml of *n*-hexane, dichloromethane, ethyl acetate, methanol and water, during 24-h, at room temperature with occasional shaking. After filtration, the extracts were concentrated under reduced pressure at 40-45 °C and stored at 4 °C until use.

Phytochemical screening

Phytochemical analysis was carried out through thin layer chromatography on silica gel plates developed with appropriated mixtures of eluents. Spots were revealed with the following spray-reagents: Dragendorff reagent for alkaloids, anisal-

Table 1 – *Lavandula taxa*, abbreviations, collection places and voucher numbers.

<i>Taxa</i> / Abbreviation	Collection place	Voucher number
<i>L. stoechas</i> subsp. <i>luisieri</i> / Lav 69	Herd. Ribeira Abaixo, Grândola	LISU 236672
* <i>L. stoechas</i> subsp. <i>luisieri</i> / Lav 74	Herd. Ribeira Abaixo, Grândola	LISU 236673
<i>L. stoechas</i> subsp. <i>luisieri</i> / Lav CB	Barragem Castelo de Bode	LISI 163/ 2010
* <i>L. pedunculata</i> / Lav Esc	Mata Nac. Escaroupim	LISI 164/ 2010
<i>L. pedunculata</i> / Lav 1	Idanha-a-Nova, Salvat. Extremo	LISI 2000/ 2009
<i>L. pedunculata</i> / Lav 2	Idanha-a-Nova, Segura	LISI 2001/ 2009

* *taxa* used in the phytochemical study

dehyde-sulfuric acid reagent for terpenes, Natural Products-Polyethylene Glycol (NEU) reagent for flavonoids and Fast Blue salt reagent for phenolic compounds, prepared according to Wagner and Blader (1996). Results were displayed semi-quantitatively in a range between absence (-) and strongly present (+++).

Light microscopy (LM)

Samples of fresh, fixed and paraffin embedded material were used. The fresh material was hand-sectioned transversely and paradermally clarified with sodium hypochlorite and washed in distilled water. Fixation was achieved in a 2.5% glutaraldehyde solution in a 0.1 M sodium phosphate buffer, pH 7.2, for 5 h at 4°C and dehydrated in a graded ethanol series (Hayat, 1981). Some fixed material was embedded in paraffin (Ruzin, 1999) and tissue blocks were sectioned at 10-12 µm using a Leitz 1512 Minot microtome.

A Nikon Labophot 2 microscope was used and images were obtained with a Nikon FX-35W camera. Characters were observed on fifteen different observations per specimen.

Scanning electron microscopy (SEM)

Plant material was fixed as above, dried with liquid CO₂ at a critical point, on a Critical Point Polaron BioRad E3500, and coated with gold on a Jeol JFC-1200. Samples were examined at 15 kV with a Jeol JSM-5220 LV scanning electron microscope. Measurements and counts were achieved on SEM micrographs obtained by computer assisted image analysis.

Histochemistry

The following histochemical tests were carried out: (a) Sudan Red for total lipids (Johansen, 1940); (b) Nile Blue procedure for neutral and acidic lipids (Jensen, 1962); (c) Copper acetate / Rubeanic acid for fatty acids (Ganter and G. Jolés, 1969); (d) Nadi reagent for essential oils and oleoresins detection (David and Carde, 1964); (e) 2,4-dinitrophenylhydrazine for terpenes with a carbonyl group (Ganter and G. Jolés, 1969); (f) Antimony trichloride for steroids (Hardman and Sofora, 1972) (g) Ferric trichloride for phenols (Johansen, 1940); (h) Vanillin hydrochloric for tannins (Johansen, 1940); (i) PAS for total polysaccharides (Feder and O'Brien (1968); (j) Ruthenium Red for pectins (Jensen, 1962); (k) Tannic acid for mucilages (Pizzolato and Lillie, 1973) and

(l) Dittmar reagent for alkaloids (Johansen, 1940). Standard control procedures were carried out at the same time. Observations were done in the mentioned LM and results were semi quantitatively analysed, between negative (-) and strongly positive (+++).

Results

Phytochemical screening

The results of the phytochemical screening and the yield of the dried extracts (as w/w percentage of the starting dried material) are presented on Table 2. It is clear that all plant extracts were negative to alkaloids, all except the methanol extract of *L. pedunculata*. The extracts obtained seemed to be rich in phenolics, flavonoids and terpenes. *L. stoechas* subsp. *luisieri* extracts seemed to have a higher content and a higher variety of terpenic compounds.

Plant trichomes

L. stoechas subsp. *luisieri* and *L. pedunculata* bear an indumentum in vegetative and flowering structures that includes non-glandular and glandular trichomes random distributed on both surfaces. The different populations maintain the characters of each species, only presenting some changes in the indumentum density. Both species showed short uni- and bicellular unbranched trichomes and mainly pluricellular branched trichomes of the stellate type. In the leaf upper epidermis the density of the last type of trichomes was smaller and they were also less long. In *L. stoechas* subsp. *luisieri* their stalk was up to 52-85 mm long and in *L. pedunculata* 60-128 mm and in both *taxa* they support 3-7 arms (Fig. 1A). In the lower epidermis this type of trichomes was denser and shorter, up to 26-39 mm in *L. stoechas* subsp. *luisieri* and 27-60 mm in *L. pedunculata*. In the flowers, only the fertile bracts and the sepals showed stellate trichomes and their stalk could reach up to 300 mm long. The petals did not have this kind of hairs, only uni- and pluricellular unbranched trichomes.

Concerning the glandular trichomes found, they belong to the peltate and capitate types. The peltate trichomes had 1 basal cell supporting 1 short stalk cell which bears an head composed of 8-12 glandular cells topped by a subcuticular space where the secretion products were stored. The capitate trichomes were much more variable in their morphology and different types could be distinguished: in

Table 2 – Semi - qualitative evaluation of the extracts chemical composition.

Plants extracts (% w/w)	Identification of extracts chemical composition			
	Phenolics	Flavonoids	Terpenes	Alkaloids
<i>L. stoechas</i> subsp. <i>luisieri</i>				
<i>n</i> -hexane (0.7)	++	++	+++	-
CH ₂ Cl ₂ (2.0)	++	++	+++	-
AcOEt (1.6)	++	+	++	-
MeOH (2.5)	++	+++	++	-
H ₂ O (4.6)	++	+	++	-
<i>L. pedunculata</i>				
<i>n</i> -hexane (0.4)	+	+	+	-
CH ₂ Cl ₂ (0.7)	++	++	+++	-
AcOEt (1.6)	++	++	++	-
MeOH (3.3)	+	+	+	+
H ₂ O (2.9)	+	++	-	-

(% w/w), extract weight / dried powdered plant material weight percentage.
 - absence; + slightly present; ++ moderately present; +++ strongly present.

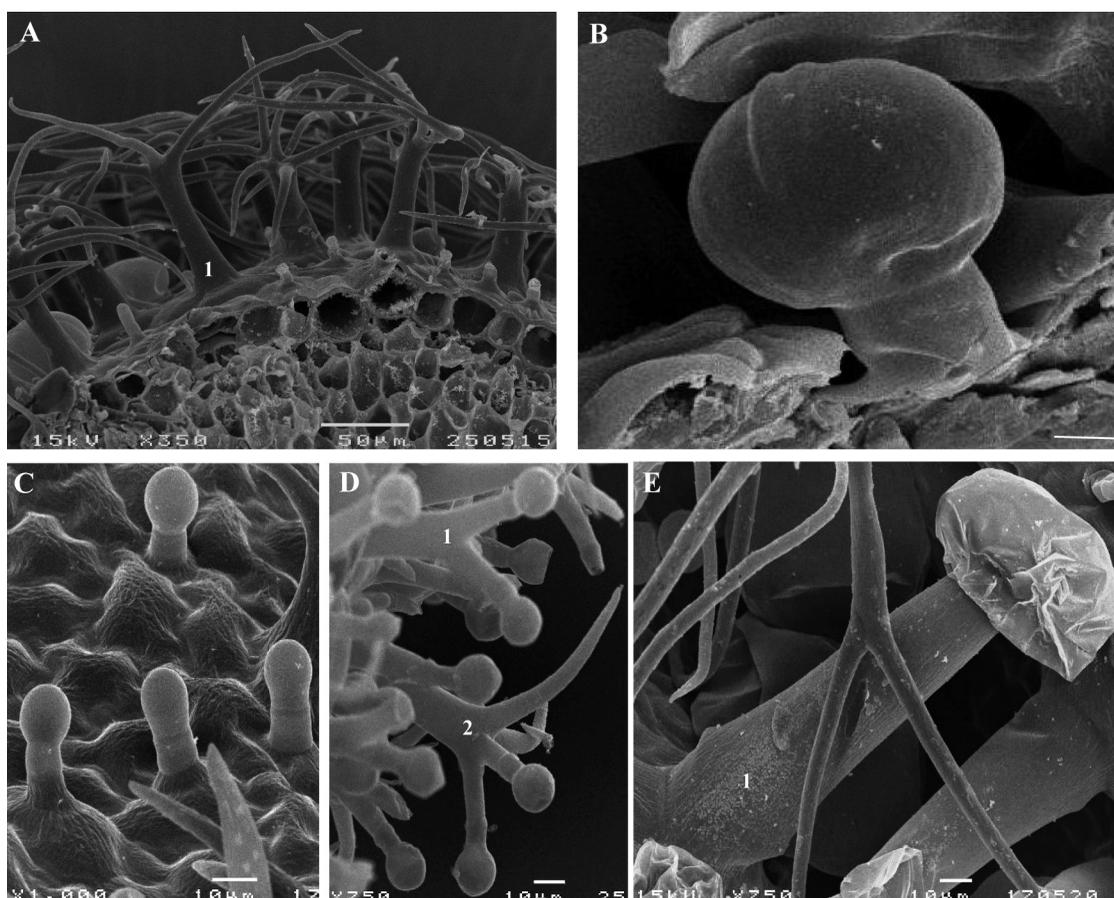


Figure 1 – A, B, C: *L. stoechas* subsp. *luisieri* SEM micrographs. A - detail of leaf transverse section showing upper epidermis with non glandular multi-cellular branched trichomes with a stalk bearing several arms, stellate type (1); B - capitulate type I (scale bar 4 mm); C - capitulate type II; D, E: *L. pedunculata* SEM micrographs: D – stellate trichome with glandular arms only (1) and stellate trichome with glandular and non-glandular arms coexisting (2); E - detail of sepal upper epidermis showing glandular trichome, with a broad stalk (1).

L. stoechas subsp. *luisieri*, capitate type I with 1 basal cell, 1 stalk cell and a head with 1-2 broad cells (Fig. 1B); capitate type II with 1 conical basal cell, 1-2 stalk cells and a round apical head (Fig. 1C). In the fertile bracts and sepals of *L. pedunculata* and in much smaller number in the leaves, the variety of secretor hairs was bigger: besides the peltate and capitate type I and II just described, there were also capitate type III trichomes. These showed 1 conical basal cell, 2-5 elongated stalk cells and a round head cell that might become cup-shaped at maturity (Fig. 3B; 3D). In this species besides the previous types of non-glandular and glandular trichomes, there were found other kinds of glandular hairs: stellate trichomes with 2-4 arms, similar in their measures and general structure to the non-glandular previous described, but their arms support glandular cells at the end, or these coexist with non-glandular end cells (Fig. 1D). In *L. pedunculata* we have found yet another type of secretor trichome, also distributed mainly in the fertile bracts and in the sepals, even if outnumbered. They showed a long and broad stalk, consisting of 2 cells, one conical elongated and another much smaller, similar to a ring, which supports a pluricellular head, and whose cells were covered by a cuticle (Fig. 1E). The petals of *L. pedunculata* did not show any of these kinds of trichomes.

Histochemistry

The results of the histochemical tests for the various types of glandular trichomes found in leaves and flowers of both *taxa* are shown in Table 3. The secreted compounds in the peltate trichomes were accumulated beneath the cuticular sheet which at maturity lifts to form a large sub-cuticular chamber for the secretory materials storage. Our observations did not reveal any pores or cracks through which the secretory material could exude. In *L. stoechas* subsp. *luisieri* specimens peltate trichomes stained slightly positive with Sudan Red and Rubeanic acid, stained positive with Nile Blue (Fig. 2A) and 2,4 - Dinitrophenilhidrazine (Fig. 2B), stained strongly positive with Nadi reagent and Antimony trichloride and stained negative with Vanillin hydrochloric, Ferric trichloride, PAS, Ruthenium Red and Dittmar reagent and the result was inconclusive with Tannic acid. In this subspecies the secretions of capitate trichomes, type I and II, found in the leaves are similar, the only difference is the staining intensity: they stained positive with Sudan Red (Fig. 2C), Nile Blue, Rubeanic acid (Fig. 2D), Antimony trichloride and Ferric trichloride, and stained negative with the remaining tests. In the flowers of *L. stoechas* subsp. *luisieri* specimens peltate and capi-

Table 3 – Histochemical tests for the indumentum of the *Lavandula taxa* under study.

Histochemical Test	Peltate		Cap I		Cap II		Cap III	Others
	<i>L. stoech.</i>	<i>L. pedunc.</i>	<i>L. stoech.</i>	<i>L. pedunc.</i>	<i>L. stoech.</i>	<i>L. pedunc.</i>	<i>L. pedunc.</i>	<i>L. pedunc.</i>
Sudan Red	+	+	++	++	++	++	+++	+++
Nile Blue	++	-	++	+	++	++	+	+
Cu Acet./Rubeanic Ac.	+	+	++	++	++	++	++	++
Nadi Reagent	+++	+++	-	+	-	+	+++	+++
2,4-Dinitrophenilhidraz.	++	+	-	+	-	++	++	++
Antimony trichloride	+++	+	++	++	+	+	-	-
Ferric trichloride	-	-	++	+	++	+	++	++
Vanillin hydrochloric	-	-	-	-	-	-	-	-
PAS	-**	+	-**	+	-**	+	+	++
Ruthenium Red	-**	+	-**	+	-**	+	++	++
Tanic acid	?	+	-**	-	-**	+	-	-
Dittmar Reagent	-	-	-	-	-	+	+	+

L. stoech is *L. stoechas* subsp. *luisieri*; *L. pedunc* is *L. pedunculata*. Cap I is capitate type I; Cap II is capitate type II; Cap III is capitate type III; - negative; + slightly positive; ++ positive; +++ strongly positive; ? inconclusive.

*The head cell stained slightly positive for neutral lipids (soft pink color); in this test the other results are positive to acidic lipids (blue color).



Figure 2 – A, B, C, D: *L. stoechas* subsp. *luisieri* histochemical tests, LM micrographs. A - peltate trichome secretion Nile Blue positive for acidic lipids (scale bar 7 μm); B - glandular peltate trichome secretion 2,4-Dinitrophenylhydrazide positive for terpenes with a carbonil group (scale bar 10 μm); C - capitate type II trichome secretion Sudan Red positive for total lipids and non glandular stellate type negative (scale bar 5 μm); D - capitate type I trichome secretion copper acetate / Rubeanic acid positive for fatty acids (scale bar 0.9 μm).

tate trichomes stained slightly positive with PAS, Ruthenium Red and Tannic acid.

In *L. pedunculata* the histochemical tests concerning the peltate trichomes, from the leaves and the flowers, are negative with Nile Blue, Ferric trichloride, Vanillin, Dittmar Reagent and they are positive, although some intensity differences, with the remaining tests, including with Tannic acid (Fig. 3A). The tests of all types of capitate trichomes are also similar on their positive results, although some intensity differences (Fig. 3B and Fig. 3C). With the Nile Blue test the capitate type III trichome basal cell stained positive for acidic lipids and the head cell stained slightly positive, a soft pink color, for neutral lipids (Fig. 3D).

The results of the histochemical tests of the other kinds of trichomes found in *L. pedunculata*, stellate

trichomes with glandular and non-glandular arms or only with glandular arms and in the other capitate type with a long and broad stalk, are similar to those found in capitate type III trichomes. The mode of secretion and accumulation of the secreted compounds in these hairs, including in the capitate type III, was beneath the cuticular sheet in a subcuticular chamber (Fig. 4A and Fig. 4B).

Discussion

The phytochemical analysis in the several extracts of the studied plants, points to a similar content of phenolics and flavonoids but the terpenic content is different. In *L. stoechas* subsp. *luisieri* extracts the number of terpenic compounds was higher

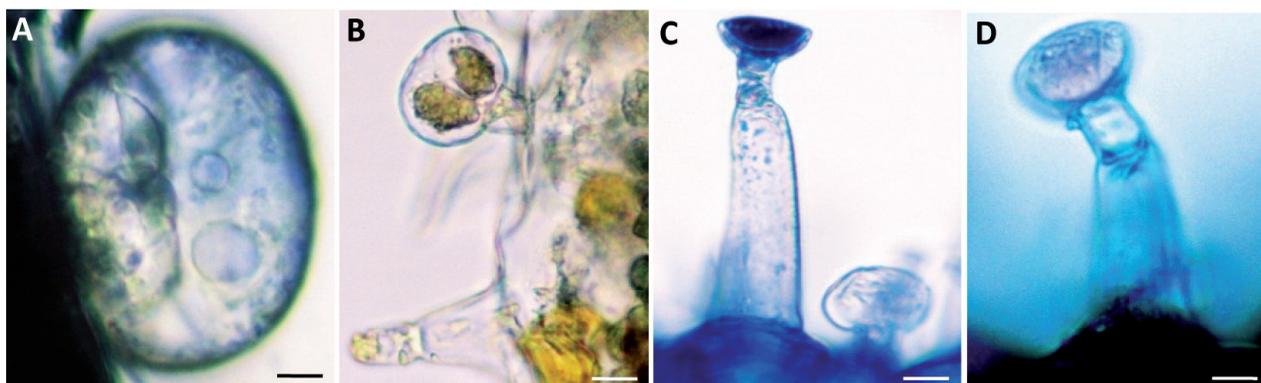


Figure 3 – A, B, C, D: *L. pedunculata* histochemical tests, LM micrographs. A - peltate trichome secretion Tannic acid positive for mucilages (scale bar 7 μm); B - capitate type III and capitate type I trichomes secretion Nadi Reagent strongly and slightly positive, respectively, for essential oils (scale bar 2,8 μm); C - capitate type II and capitate type I trichomes secretion Antimony trichloride slightly positive and positive, respectively, for steroids (scale bar 2 μm); D - capitate type III trichome secretion Nile Blue basal cell positive for acidic lipids and head cell slightly positive for neutral lipids (soft pink colour) (scale bar 2,8 μm).

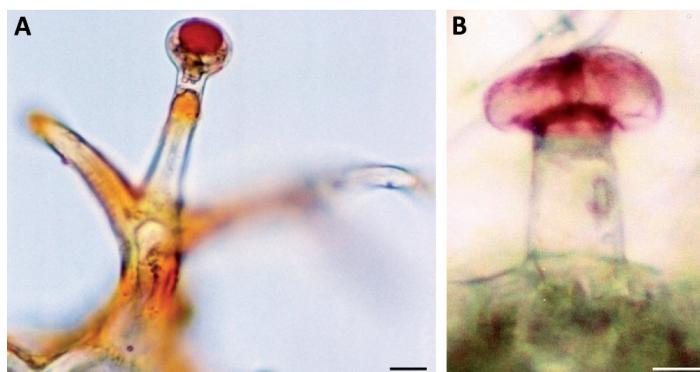


Figure 4 – *L. pedunculata* histochemical tests, LM micrographs. A - stellate multi-cellular trichome with glandular and non-glandular arms (mixed stellate type II), Sudan Red positive for total lipids (scale bar 9 μm); B – Glandular trichome with a long and broad stalk, consisting of 2 cells, one conical elongated and another much smaller, similar to a ring, which supports a pluricellular head, Ruthenium Red positive for pectins (scale bar 7 μm).

and richer. Several authors mentioned that some necrodane derivatives characterize its essential oils (Feijão, 2011; González-Coloma *et al.*, 2006; Matos *et al.*, 2009; Baldovini *et al.*, 2005; Sanz *et al.*, 2004; Garcia-Vallejo *et al.*, 1994), which seem to be absent in the remaining *Lavandula* spp. essential oils. Several authors working on Spanish material of *L. stoechas* mention the difficult differentiation of subspecies and varieties within this species due to high polymorphism, but they consider some chemotypes based only on chemical data (Garcia-Vallejo *et al.*, 1994). The essential oils of *L. pedunculata* are rich in fenchone (Zuzarte *et al.*, 2009). In a previous work with specimens harvested in our locations, Feijão (2011) also confirmed the dominance of fenchone in the chemical composition of the essential oils of the same species.

L. stoechas subsp. *luisieri* and *L. pedunculata* showed non-glandular trichomes, small uni- and bicellular trichomes and abundant stellate trichomes. These hairs have a similar distribution in the leaves fertile bracts and sepals surfaces but stellate trichomes are not found in the petals. Based on their structure, glandular trichomes are generally classified as either peltate or capitate (Werker *et al.*, 1985). Peltate trichomes do not present any diversity in their morphology; they are similar to those found in other Lamiaceae. The capitate trichomes vary greatly in structure and size. According to our observations *L. pedunculata* showed a wider diversity of secretory trichomes, such as the multicellular stellate trichomes which combine glandular and non-glandular arms. Zuzarte *et al.* (2010) also mentioned this kind of trichomes in samples of *L. pedunculata* from central north Portugal as well as in *in vitro* leaf cultures but no reference is done to

the capitate type III and to the other type of capitate trichomes, those with the long and broad stalk with the pluricellular head that we found distributed either by the leaves or by the different floral parts but once again not in the petals. This kind of glandular trichomes is mentioned by Doagey (1992) in leaves and flowers of *L. stricta* and *L. coronopifolia*, exemplars from the south of Saudi Arabia.

It should be noted that in the flowers of both *taxa* the petals showed little diversity of secretory trichomes. Only peltate hairs were seen and the greatest diversity and amount of trichomes arises in the fertile bracts and the sepals.

Data from histochemical tests showed that the metabolites accumulated in the glandular trichomes of the studied *taxa* have a distinct composition and that the secreted materials are complex in their nature. These tests contribute to a better understanding of the location of metabolites in cells and glandular structures. In the peltate hairs of *L. stoechas* subsp. *luisieri* leaves, the secreted compounds accumulated beneath the cuticle were most lipophilic, while in *L. pedunculata* leaves and flowers structures the presence of polysaccharides, mucilages and pectines were also revealed with PAS, Tannic acid and ruthenium red histochemical tests, respectively. In *L. stoechas* subsp. *luisieri* a slightly presence of these hydrophilic compounds was noticed in the fertile bracts and the sepals. The presence of these compounds in the glandular trichomes of other Lamiaceae has been reported by several authors (Werker, 1993; Corsi and Bottega, 1999; Ascensão *et al.*, 1999; Marin *et al.*, 2010; Rodrigues *et al.*, 2013) but their function is not completely explained (Werker, 1993; Corsi and Bottega, 1999).

The histochemical results indicate that capitate trichomes of *L. pedunculata* produced a much more heterogeneous secretion than those of *L. stoechas* subsp. *luisieri*. Indeed capitate trichomes seem to contain a much more complex mixture of secretory products, containing several lipophilic compounds in addition to phenols, polysaccharides and traces of alkaloids. Similar contents have also been reported in other related genus of the Lamiaceae (Serrato-Valenti *et al.*, 1997; Ascensão *et al.*, 1999; Rodrigues *et al.*, 2008; Rodrigues *et al.*, 2013). It is accepted that some of these secondary metabolites have important ecological effects and provide plants with physical and chemical protection against herbivores and pathogens (Werker *et al.*, 1985; Fahn, 1988; Harborne, 1997).

In *L. pedunculata* the other types of trichomes also have a mixture of lipophilic and hydrophilic compounds, very similar to those found in capitate type III trichomes. It is important to note that in the trichomes with glandular and non-glandular arms, both of them seem to accumulate secretions (Fig. 4A). Although their structure is quite different these trichomes seem to be more close to glandular trichomes in their secretory material and in their accumulation processes: beneath the cuticular sheet in a sub-cuticular chamber. The non-glandular hairs particularly the multi-cellular branched trichomes presumably assist glandular hairs in the mechanical defense and they are certainly also involved in preventing an excessive leaf transpiration.

Although the low specificity of some histochemical tests, they are widely used to locate metabolites in all kinds of glandular structures (Ascensão *et al.*, 1999; Bisio *et al.*, 1999; Combrink *et al.*, 2007). These tests do not allow us to identify compounds just find the main chemical groups at the cellular level. In this sense the results from Table 2 and Table 3 show some similarity between species, concerning the presence of the main chemical groups, such as the alkaloids but also the phenolics and terpenes.

In conclusion, *L. stoechas* subsp. *luisieri* and *L. pedunculata* have different types of glandular trichomes and despite the need to continue the chemical characterization of each *taxon*, our phytochemical screening and the results of the histochemical tests point to a distinct chemical composition between them.

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